

MALDI Imaging Mass Spectrometry

Technical Advances and Clinical Problem Solving

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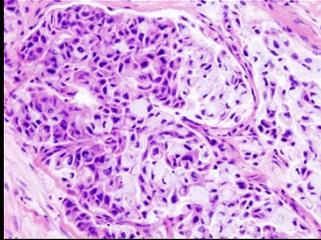
Vanderbilt University School of Medicine

March 4, 2014

Tissue Pathology

The Current Approach

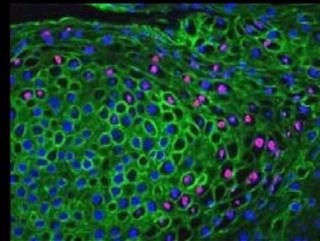
Tissue Prep



Analysis



Diagnosis



Reagents and Methods

Molecular Staining

Time and Money

The Facts

Fact: Historical approaches to tissue pathology are inadequate to answer many important clinical questions.

- Early detection
- Aggressiveness of disease
- Optimal treatment
- Drugs distribution/efficacy

Fact: Molecular testing has revolutionized the practice of medicine.

Fact: Relatively few molecular tests can be related to specific tissue pathology.



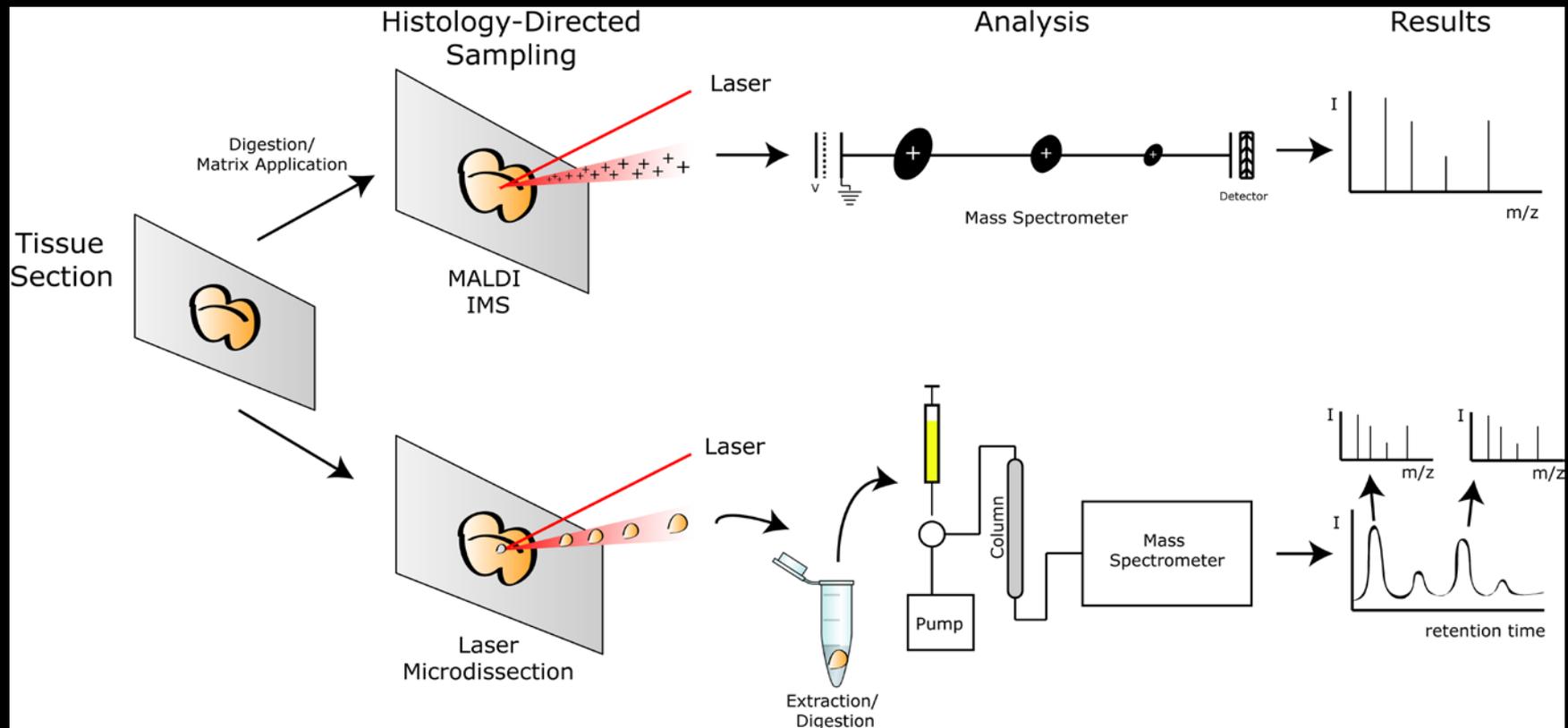
Anatomic

^ Pathology

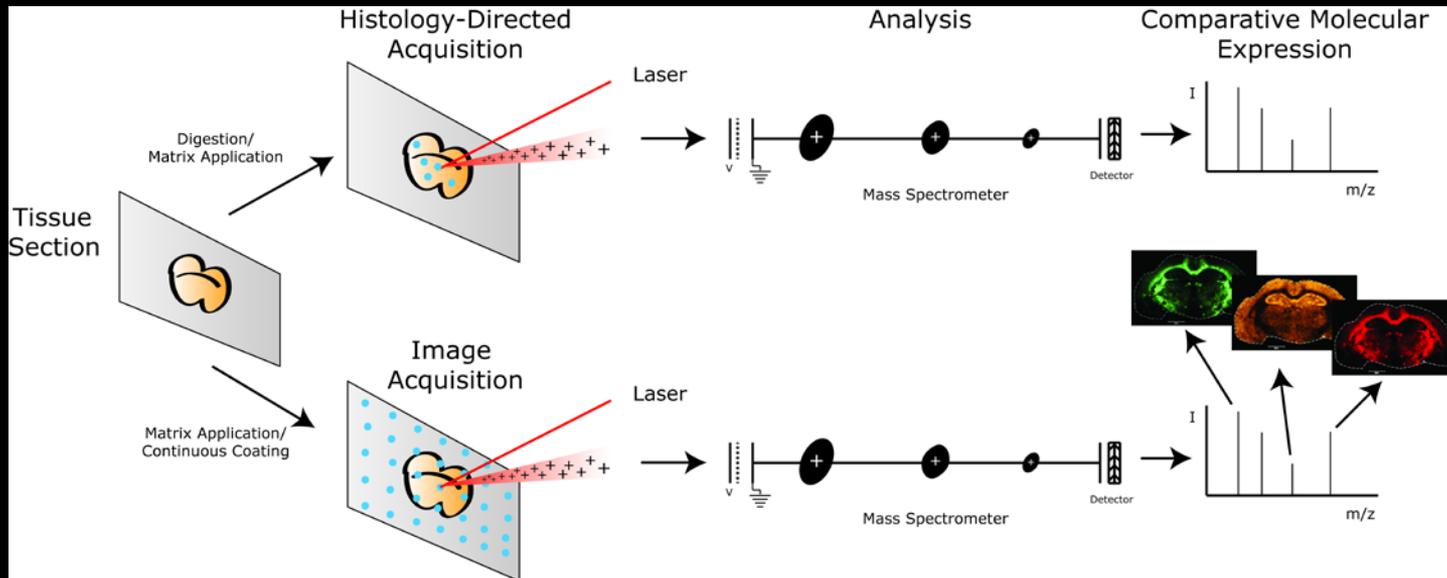
+ Mass Spectrometry

Clinical Value

Histology-Directed Analysis

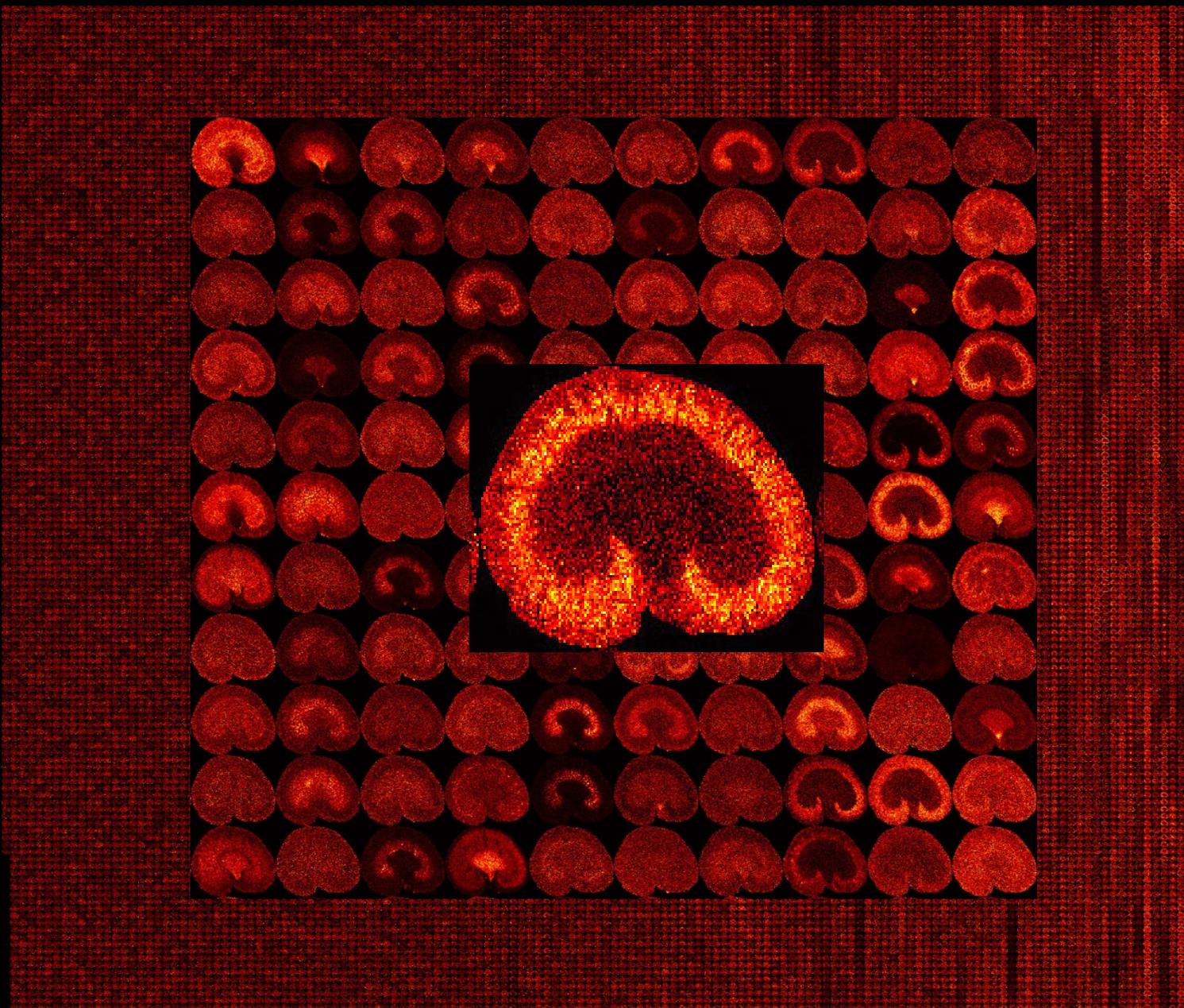


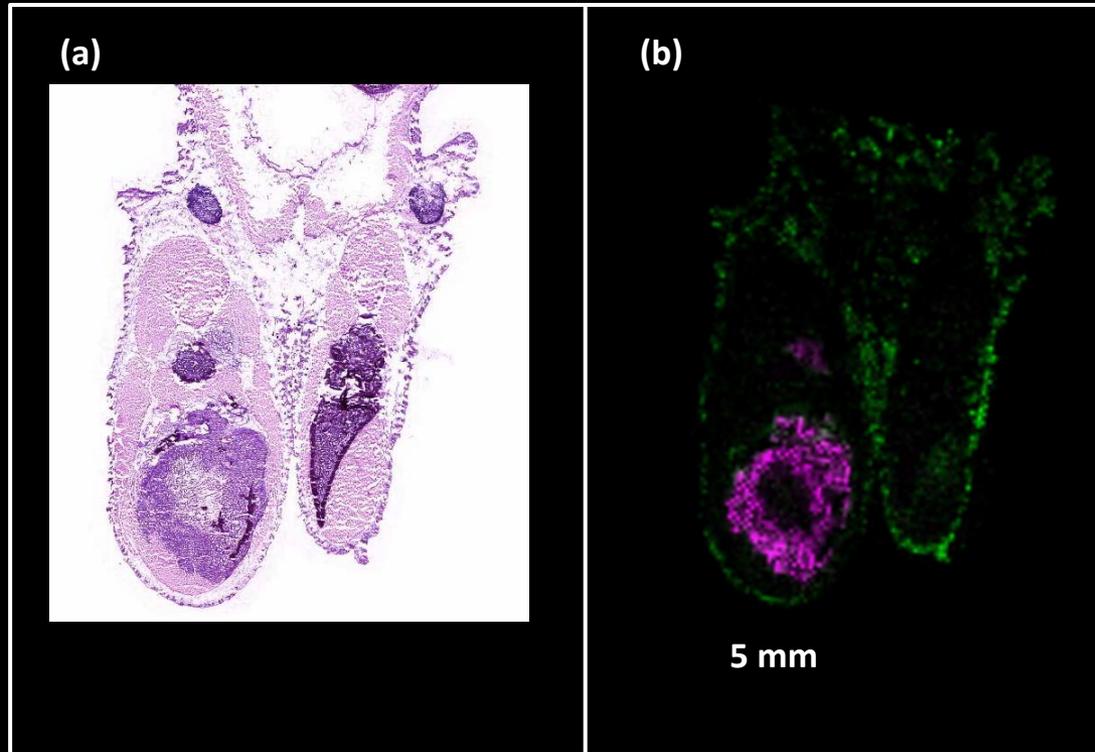
Imaging Mass Spectrometry



- Matrix applied to the tissue surface.
- Laser desorbs and ionizes molecules from the tissue surface.
- Mass spectrometer analyzes ionized molecules creating a molecular profile (fingerprint) at each position of the tissue.
- Molecular fingerprint is used for 1) disease classification and 2) analyzing molecular distribution of tissue.

Mouse Kidney – MALDI Imaging MS at 100 μm spatial resolution, 1 kHz rep rate laser, SA matrix

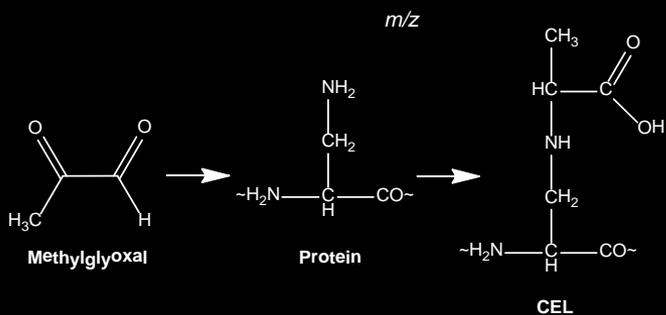
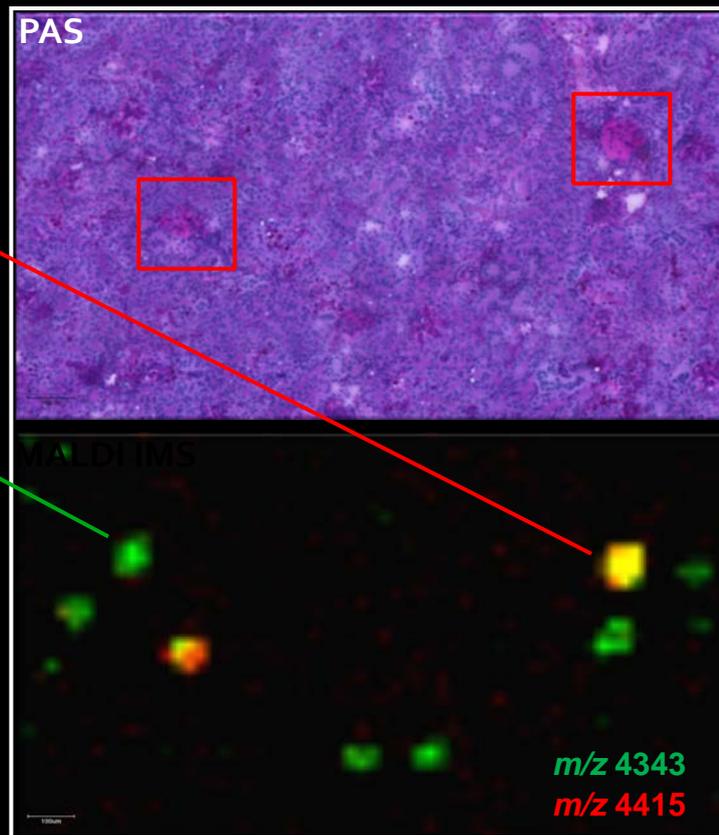
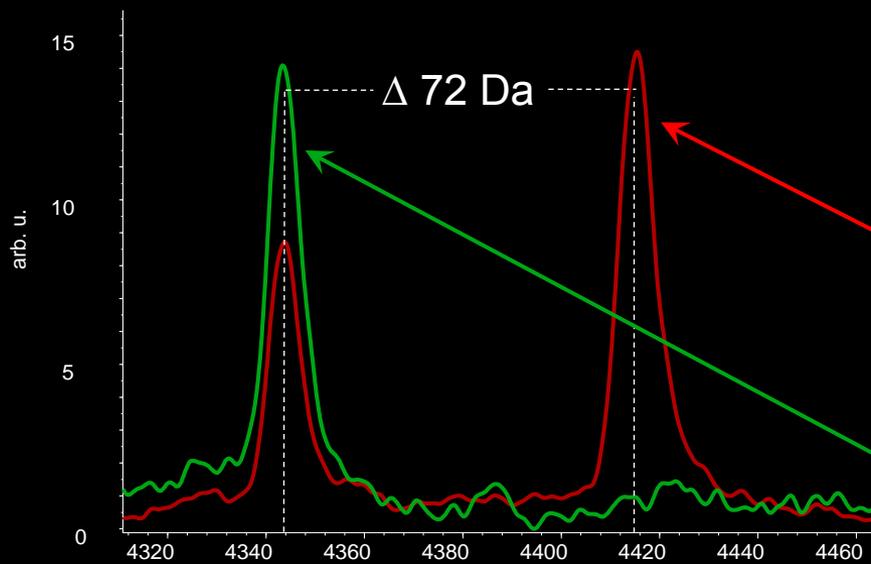




Human breast tumor cell line implanted into the tibia of a mouse.

Human calcyclin (m/z 10,090)

Mouse calcyclin (m/z 9960)



IMS Performance Criteria

Traditional Considerations

Mass Resolving Power: defined as $m/\Delta m$

Mass Accuracy: the difference between the measured mass and the calculated exact mass.

Sensitivity: specifies the overall response of the instrument for a given analyte.

Dynamic Range: detection range for the instrument (most intense/smallest detectable signal)

MSⁿ capabilities: ability to perform fragmentation experiments for analyte identification.

IMS Special Considerations

Spatial Resolution: distance between two adjacent pixels (ablated spots) on the sample surface.

Throughput: the number of scans/spectra that can be acquired per unit time.

File Size/Data Storage: Considerations of storage costs and processing practicality

Instrumental / Methodology Challenges

- Speed** - data collection too slow to be practical for routine analysis
- Sensitivity** - achieve more global coverage (fraction of analytes observed)
- Resolution** - better lateral resolution (single cell imaging)
- higher MS resolution (better resolve isoforms, PTMs)
- Mass Range** - routinely beyond 100 kd
- Identification** - *in situ* - fast, simple, accurate
- Quantitation** - reagents and methods - isotope based, relative and absolute
- Validation** - cross-lab, cross-platform reproducibility/standardization
- Availability** - single manufacturer provides entire technology 'solution'
- instruments and protocols are too complex for non-experts

Increasing Throughput

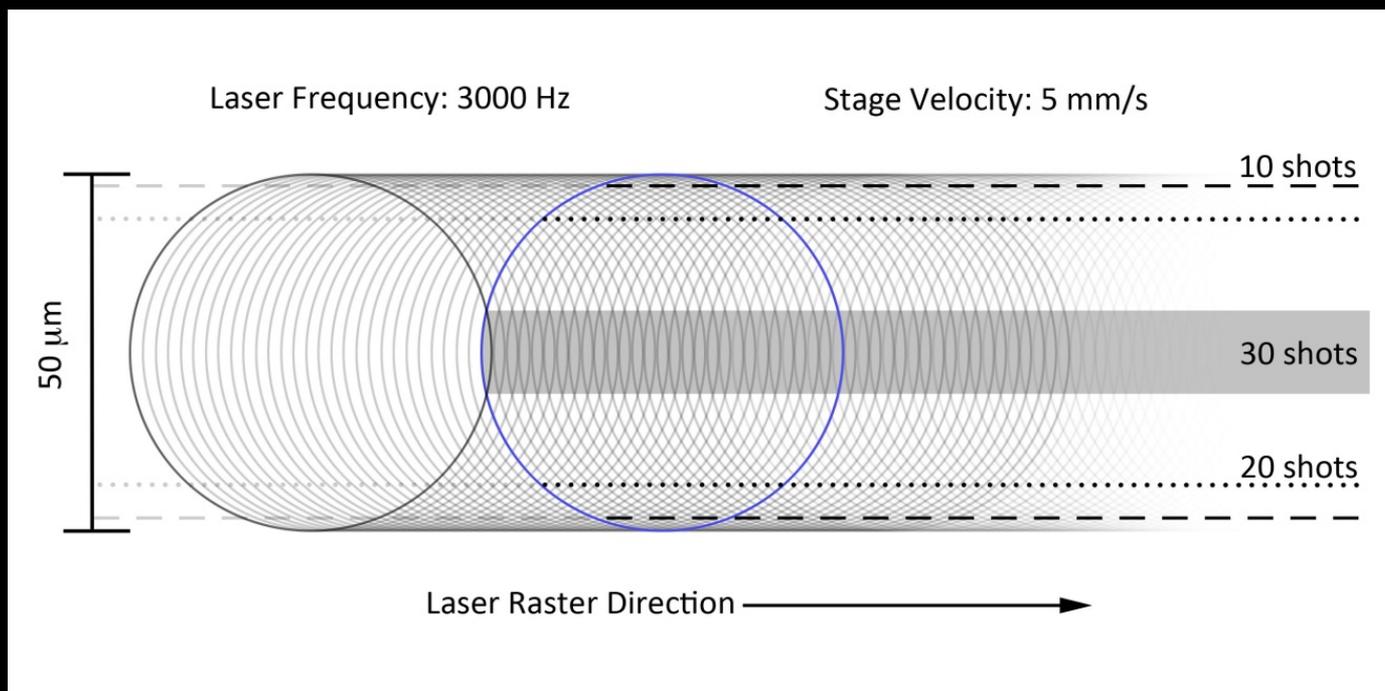
Simultof Systems

High-Speed MALDI-TOF IMS

Continuous Laser Raster Sampling

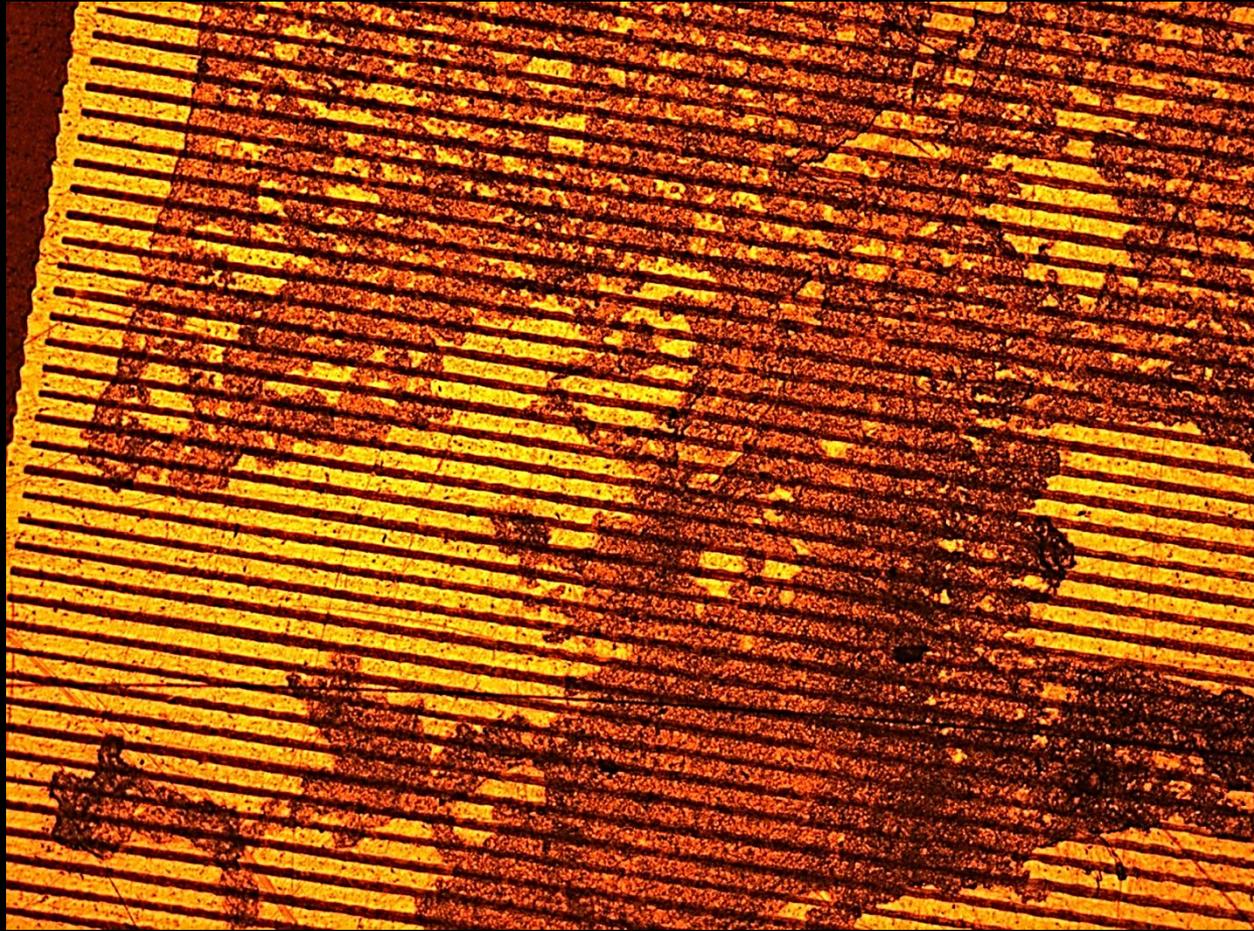
$$\text{Laser Shots/Unit Area} = f_{rep} \left(\frac{d}{v_{stage}} \right)$$

$$\text{Lateral Spatial Resolution} = H.A. \left(\frac{v_{stage}}{f_{rep}} \right)$$



High-Speed MALDI-TOF IMS

Continuous Laser Raster Sampling



High Speed MALDI TOF SimulTOF

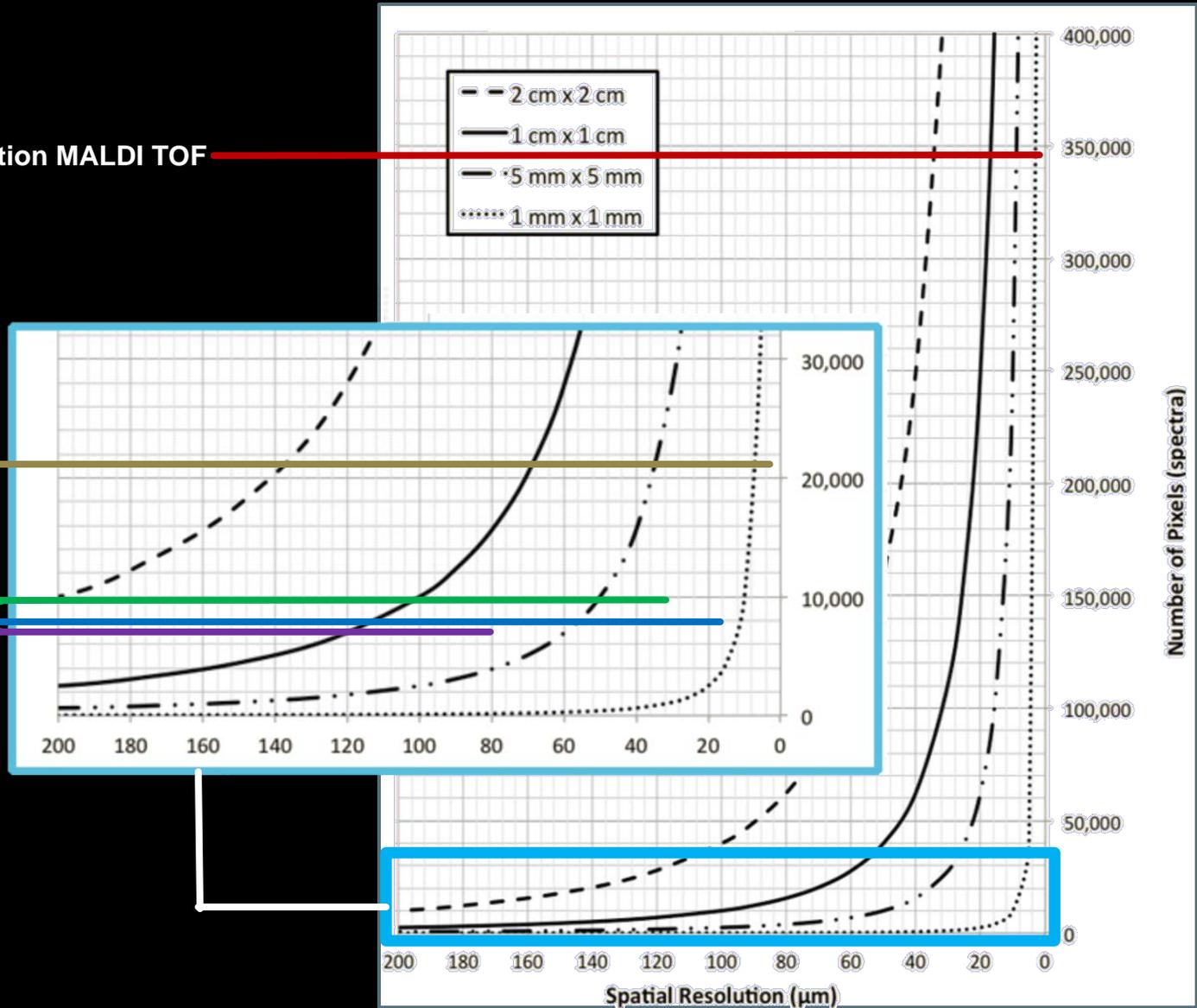
~345,000 pixels/3hr Next Generation MALDI TOF

~21,000 pixels/3hr MALDI TOF

~9,800 pixels/3hr LIT

~7,700 pixels/3hr FT-ICR

~7,200 pixels/3hr IM Q-TOF

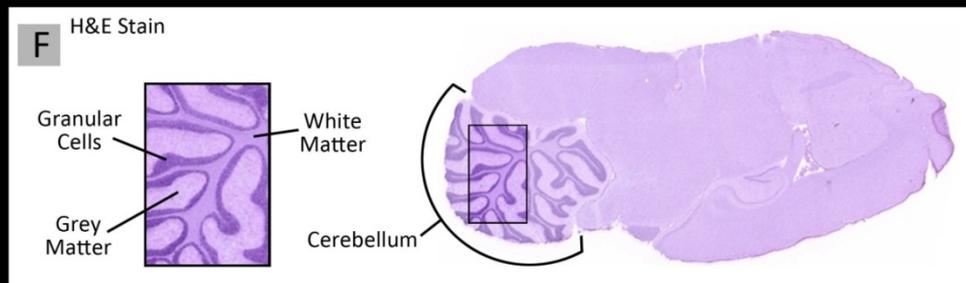
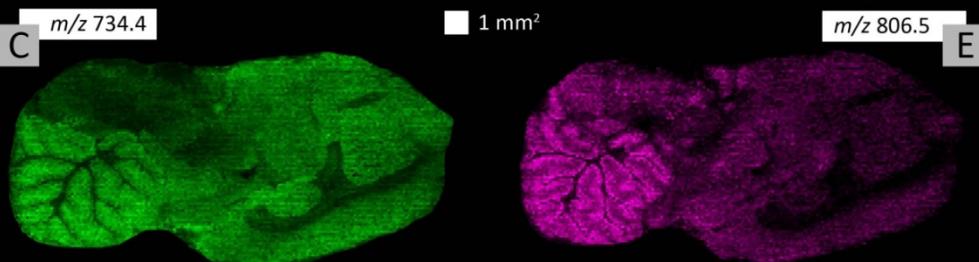
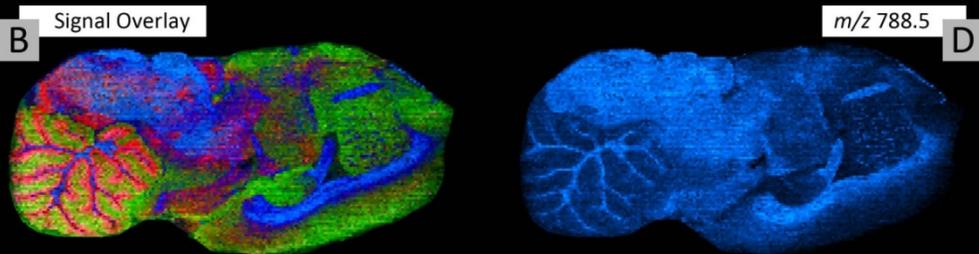
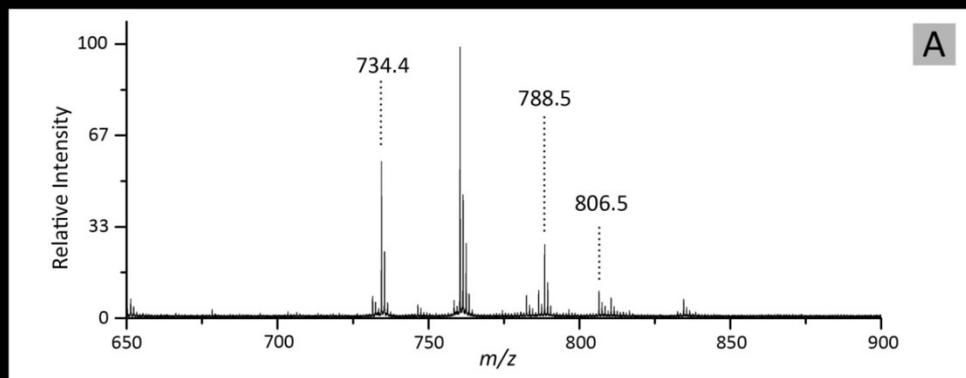


Lipid Imaging SimulTOF

Experimental Details

- **Lipids – m/z 600-1200**
- **3 kHz Laser Rep Rate**
- **Velocity: 5 mm/s**
- **Lateral Resolution: 100 μ m**
- **Vertical Step: 100 μ m**
- **Total Pixels: ~19,000**
- **Effectively: ~30 Spectra/s**
- **Total File size: 4.33 GB**

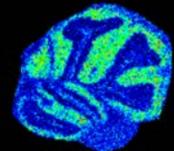
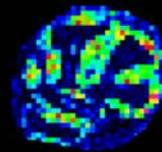
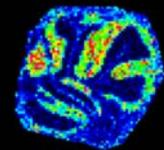
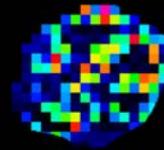
- **Time: ~10 min**



Increasing Spatial Resolution

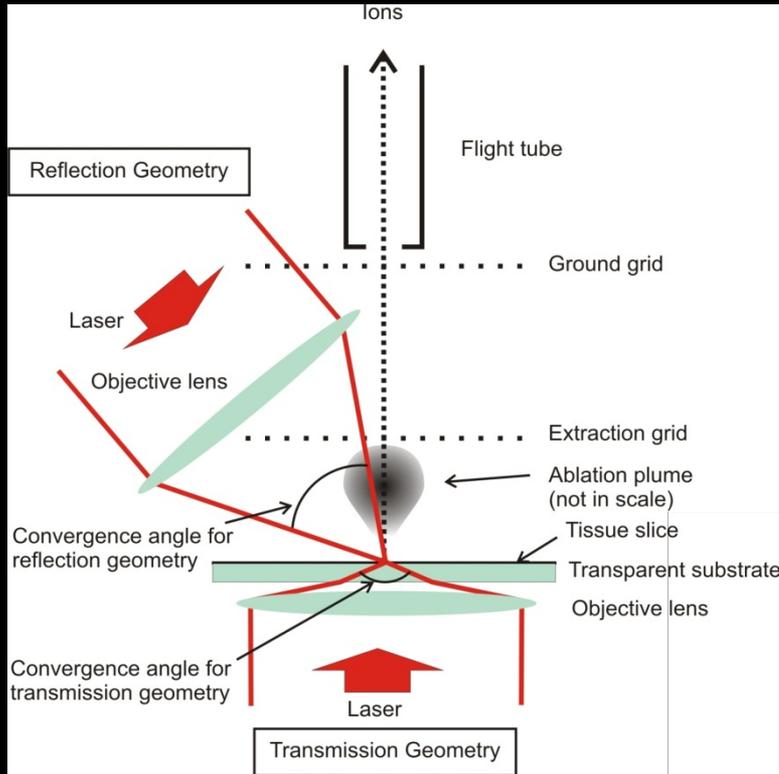
Spatial Resolution

- Image resolution defines the nature of molecular information that can be derived from an IMS experiment.
- The price of higher resolution can be significant time, effort, and money.
- Some biological questions can only be answered by high resolution imaging.



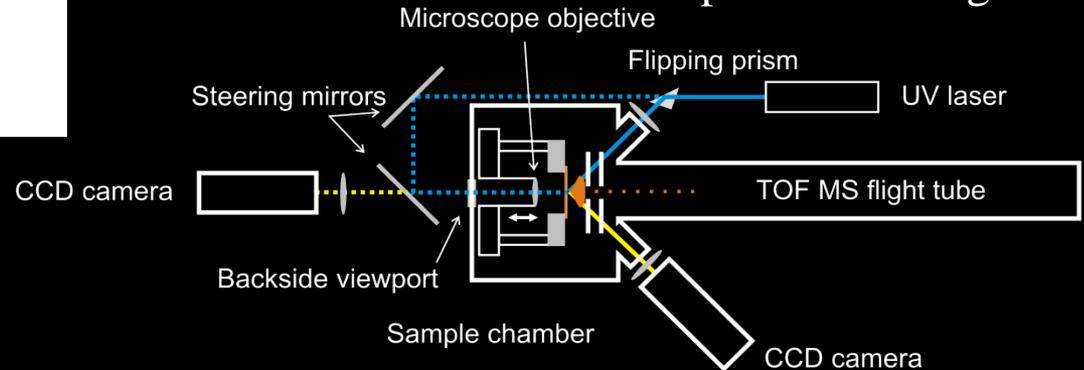
Ion image of mouse cerebellum (m/z 6765) at spatial resolutions of 200 μm , 100 μm , 50 μm , and 25 μm .

Transmission vs. reflection geometry



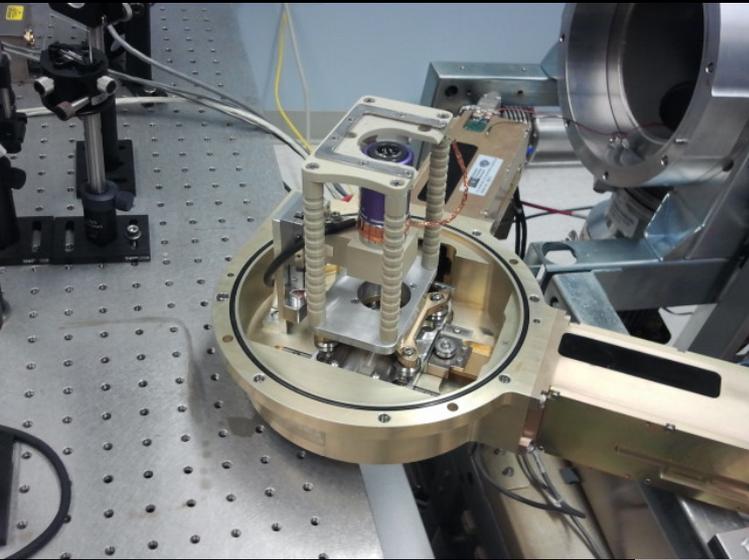
Why transmission geometry?

- Better access to target, permitting shorter working distance lenses.
- On-axis sample visualization permits better accuracy and better image quality at higher optical magnification in the instrument.
- Backside illumination may favor ion formation for matrix pre-coated targets.



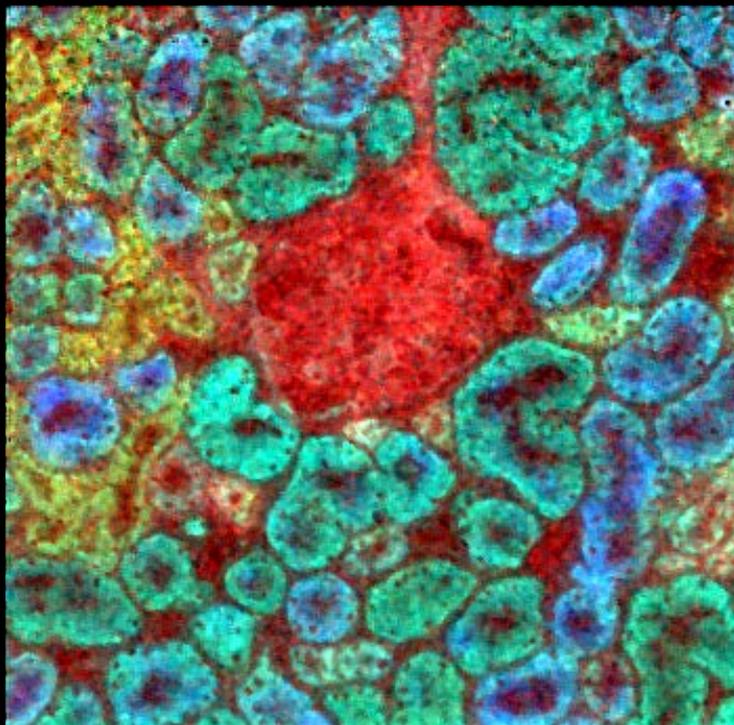
Transmission geometry Prototype

Modified AB 4700

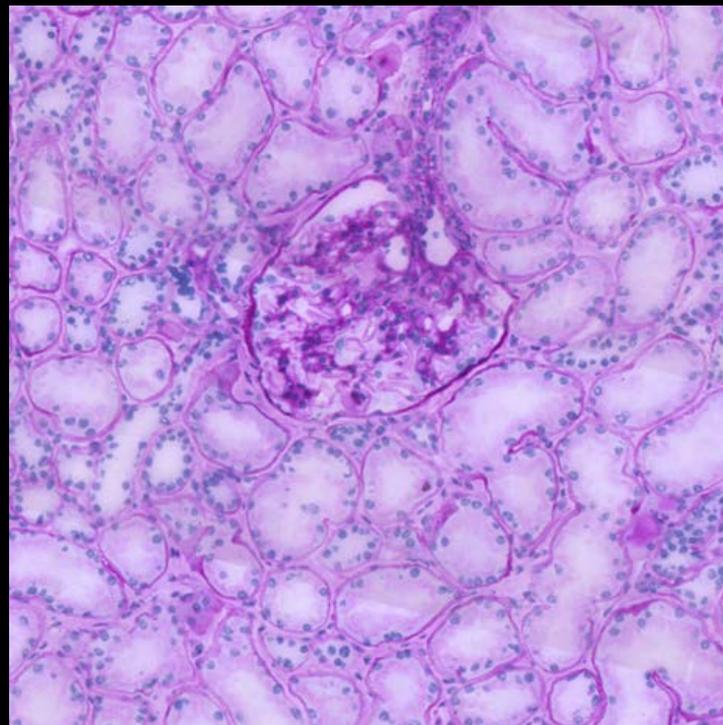


High Spatial Resolution Imaging using Transmission Geometry MALDI MS

Human Glomerulus



Imaging MS
(2 μm spatial resolution)



PAS Stain
(serial section)

Ion Overlay with Tentative IDs

Red = m/z 750; PE(P-38:4)

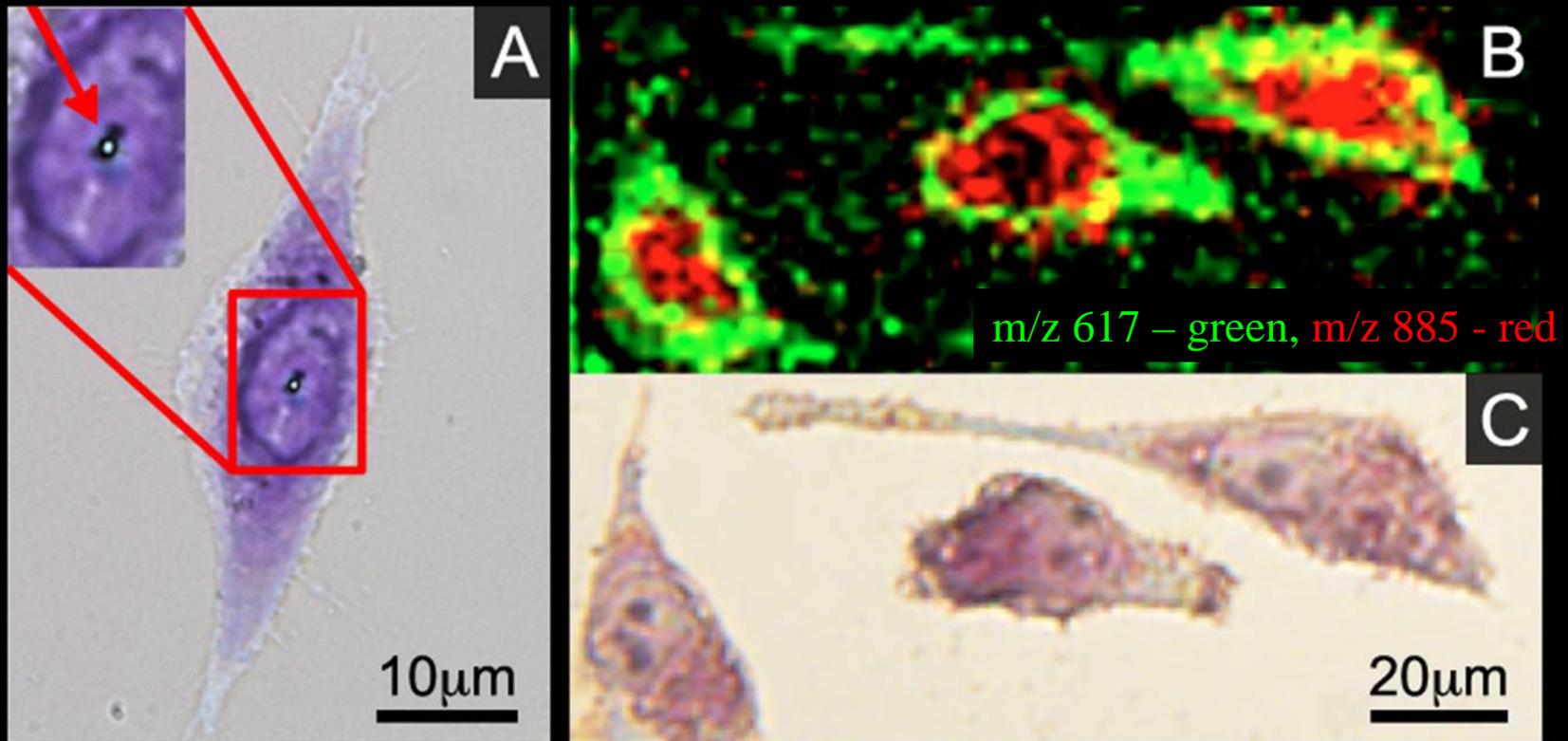
Yellow = m/z 863; PI(36:1)

Green = m/z 885; PI(38:4)

Blue = m/z 1052; SM3(d18:1/24:0)

Transmission Geometry AB4700 instrument

single cell imaging

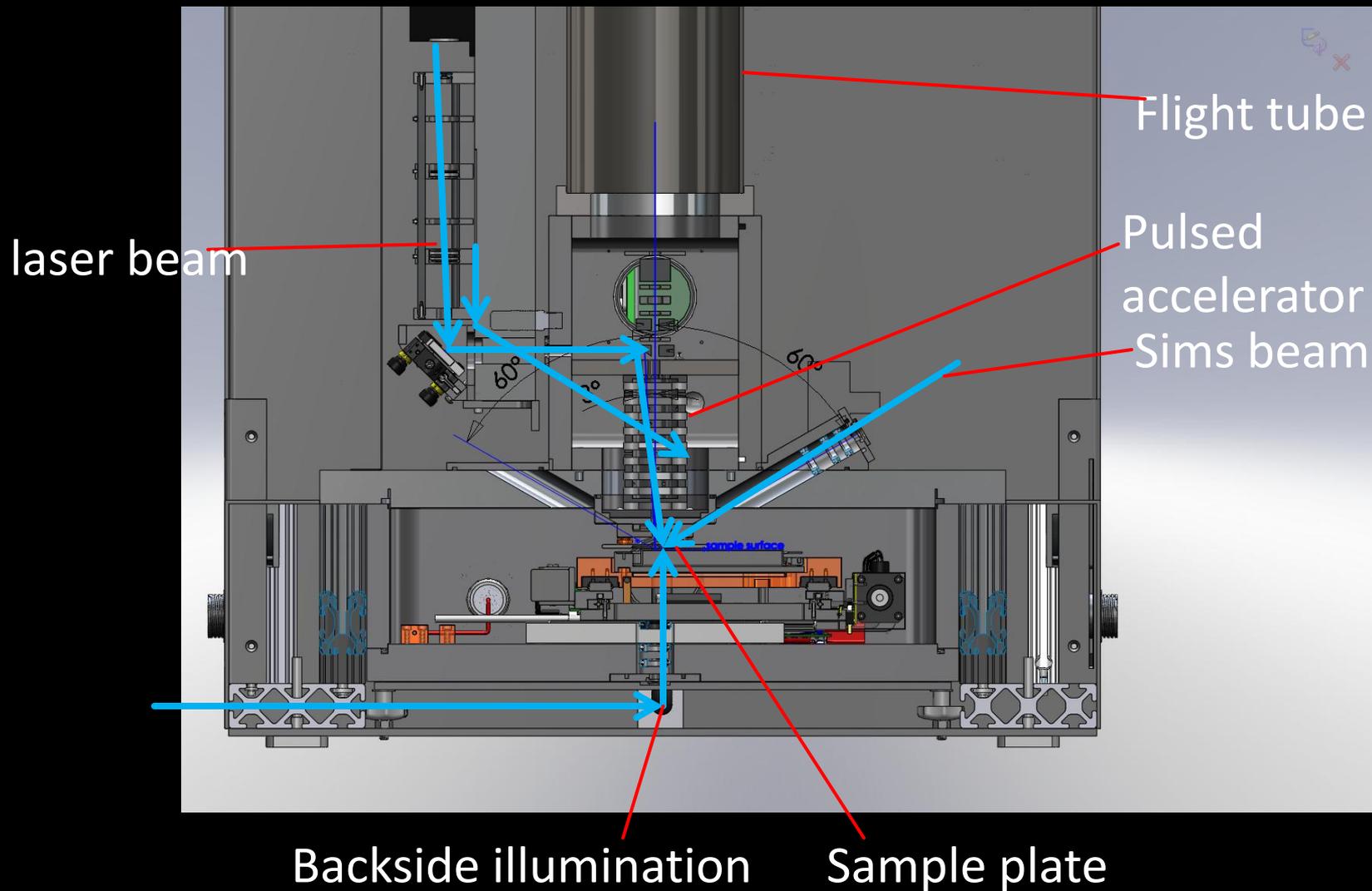


- A. Optical microscopy image of laser ablated spot in a nucleus area of a single HEK 293 cell in point-and-shoot mode
- B. MS images of single HeLa cells at raster step 2 μm.
- C. Optical microscopy image of the same HeLa cells before the MS imaging raster scan.

Angle	min spot(μm)
normal	25
3 deg	2.5
180 deg	1

SimuTOF Systems

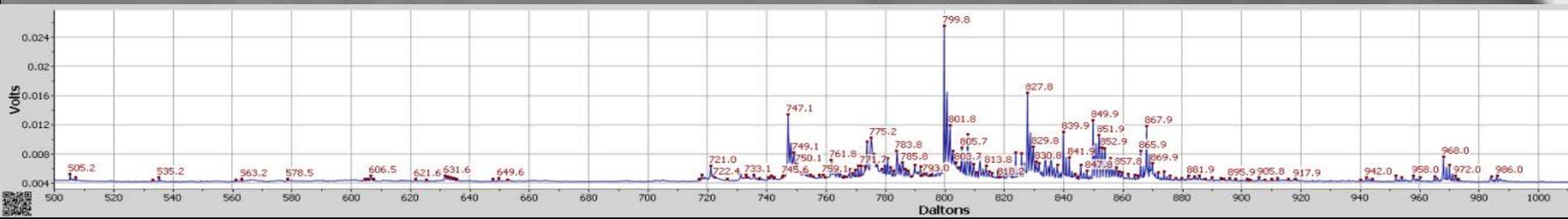
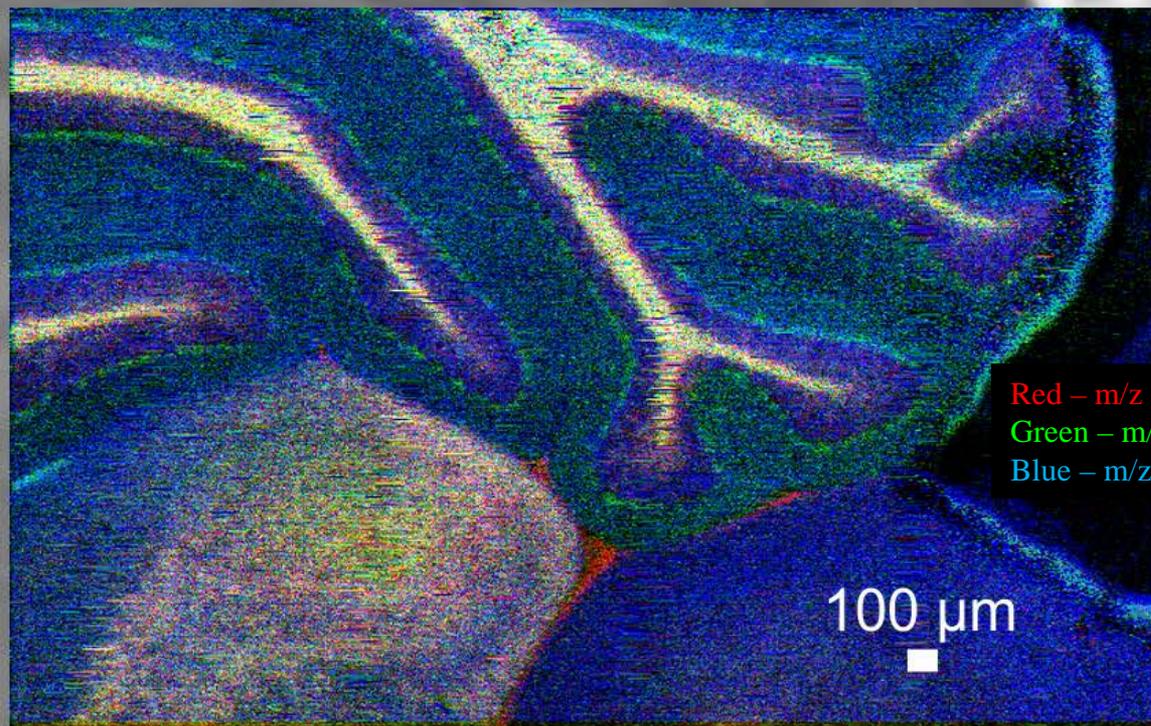
Multiport Sample Interrogation



SimulTOF instrument in transmission geometry

5 μm pixel size, $\sim 300,000$ spectra

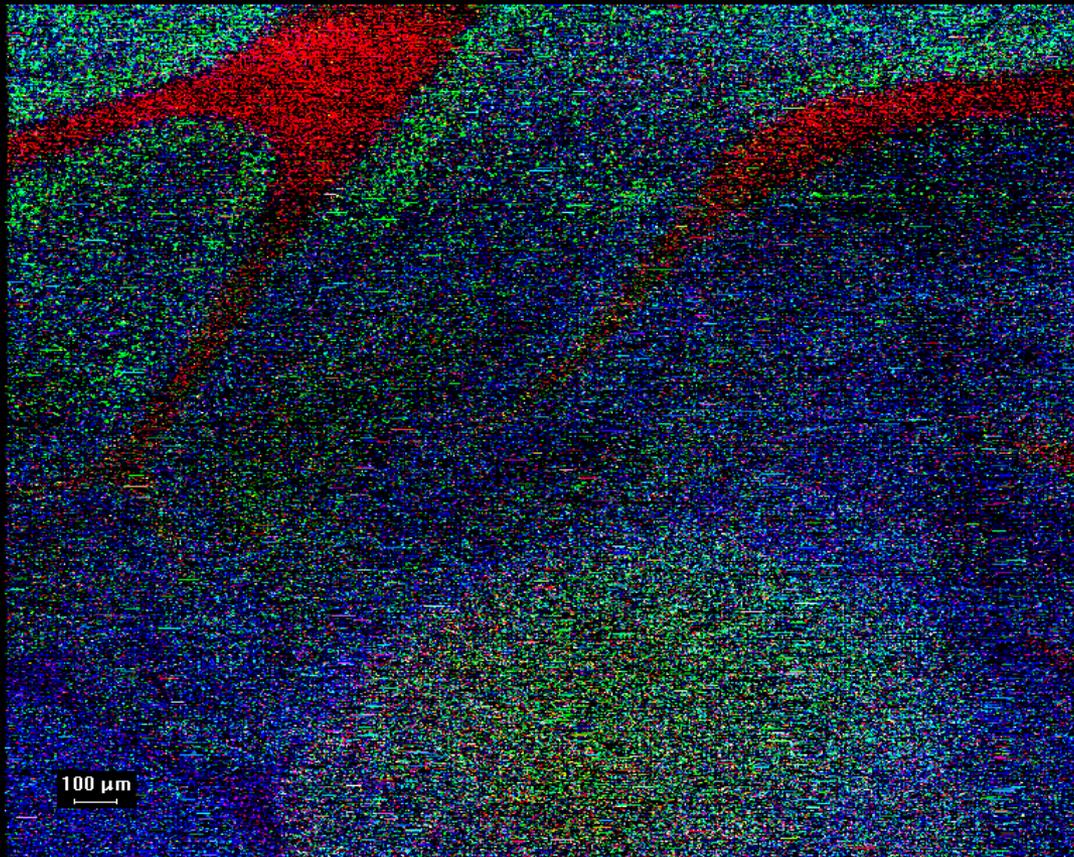
Mouse cerebellum, DHA matrix / positive mode



SimulTOF instrument in transmission geometry

2 μm pixel size, $\sim 500,000$ spectra

Mouse cerebellum, DHA matrix / positive mode



Red – m/z 827
Green – m/z 773
Blue – m/z 869

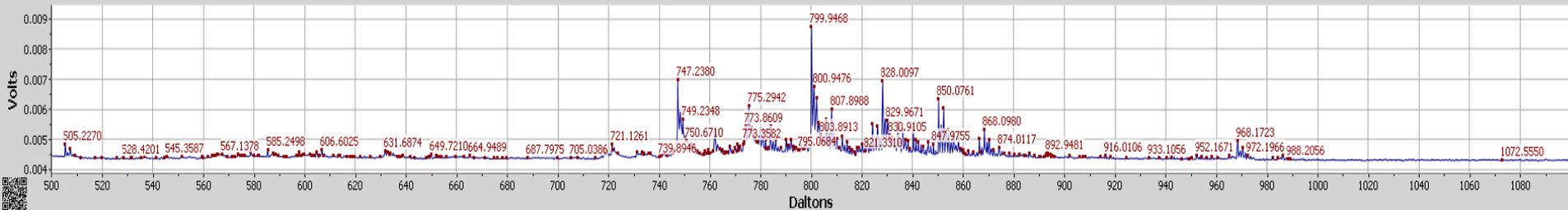
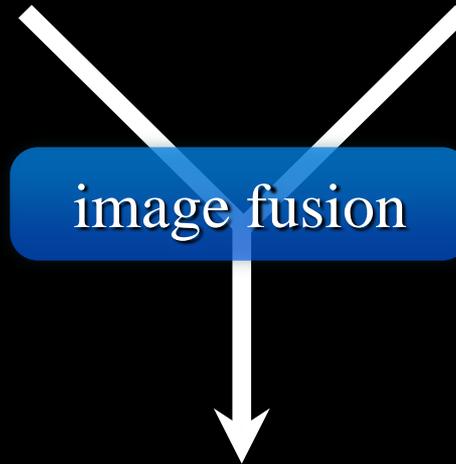


Image Fusion

What?

imaging modality 1

imaging modality 2



combined imaging modality
(never physically measured)

Image Fusion

Why?

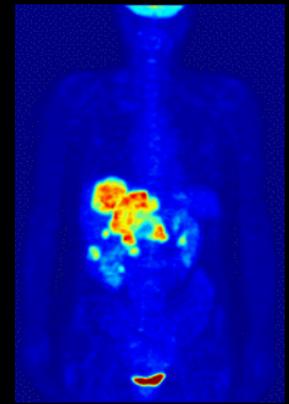
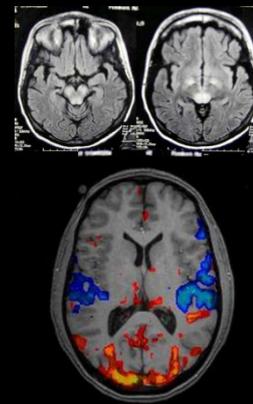
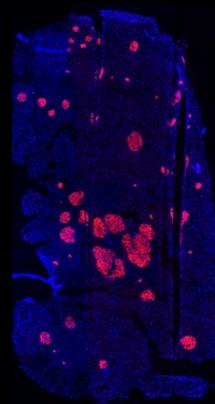
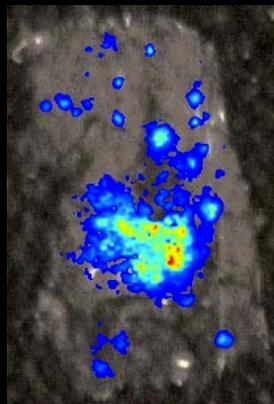
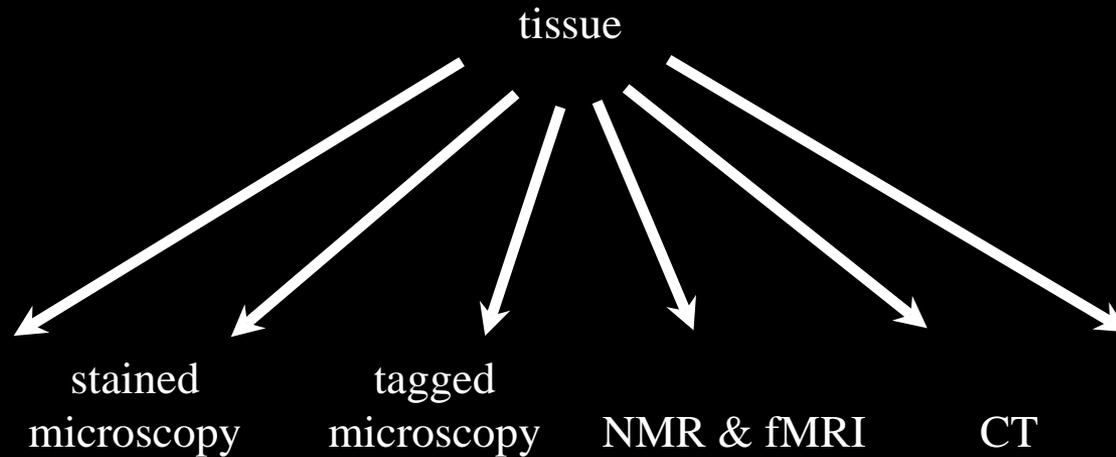


Image Fusion

Example of sharpening

pan-chromatic image

- high spatial resolution
- little info per pixel



multispectral image

- low spatial resolution
- a lot of info per pixel



fused image

- high spatial resolution
- a lot of info per pixel

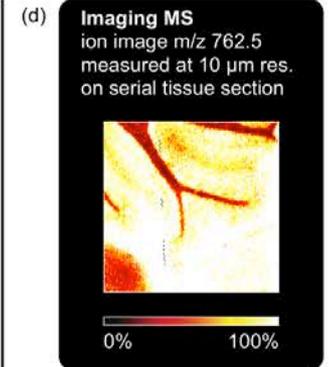
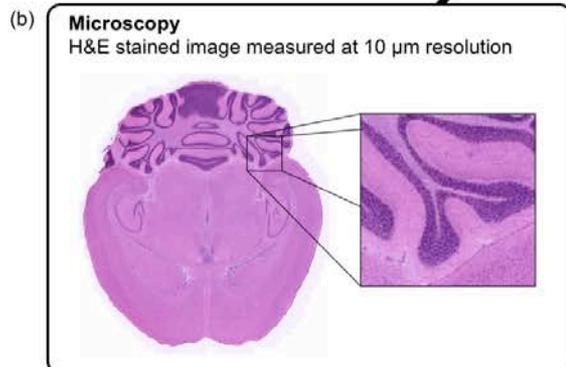
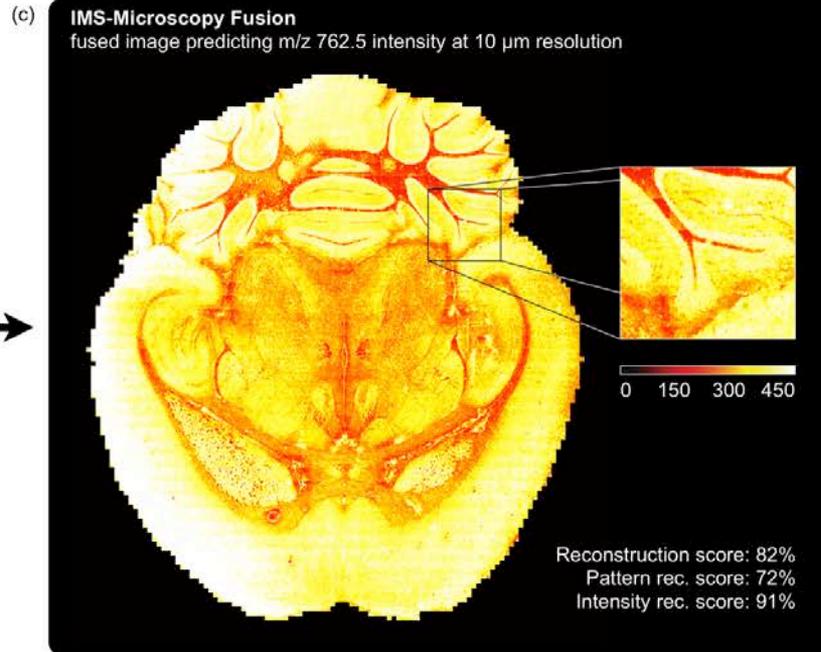
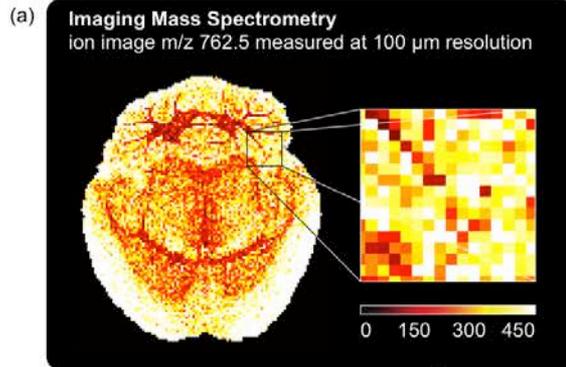


Image Source: ©2004 DigitalGlobe, Inc. All RIGHTS RESERVED

Image Fusion

Improved prediction and benchmarking

Prediction versus measurement benchmark.



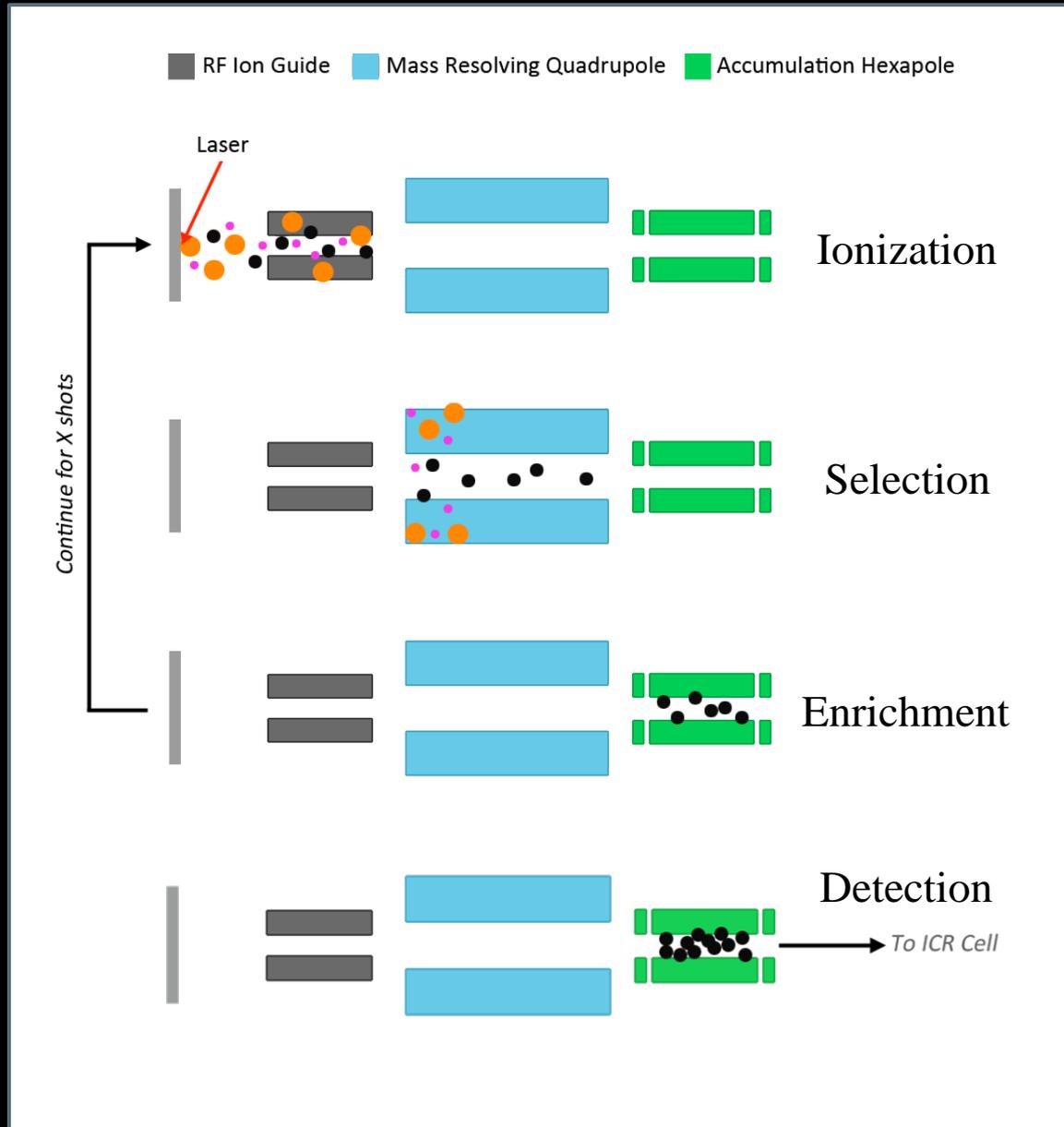
Increasing Sensitivity & Specificity

High Dynamic Range FT-ICR Imaging Mass Spectrometry

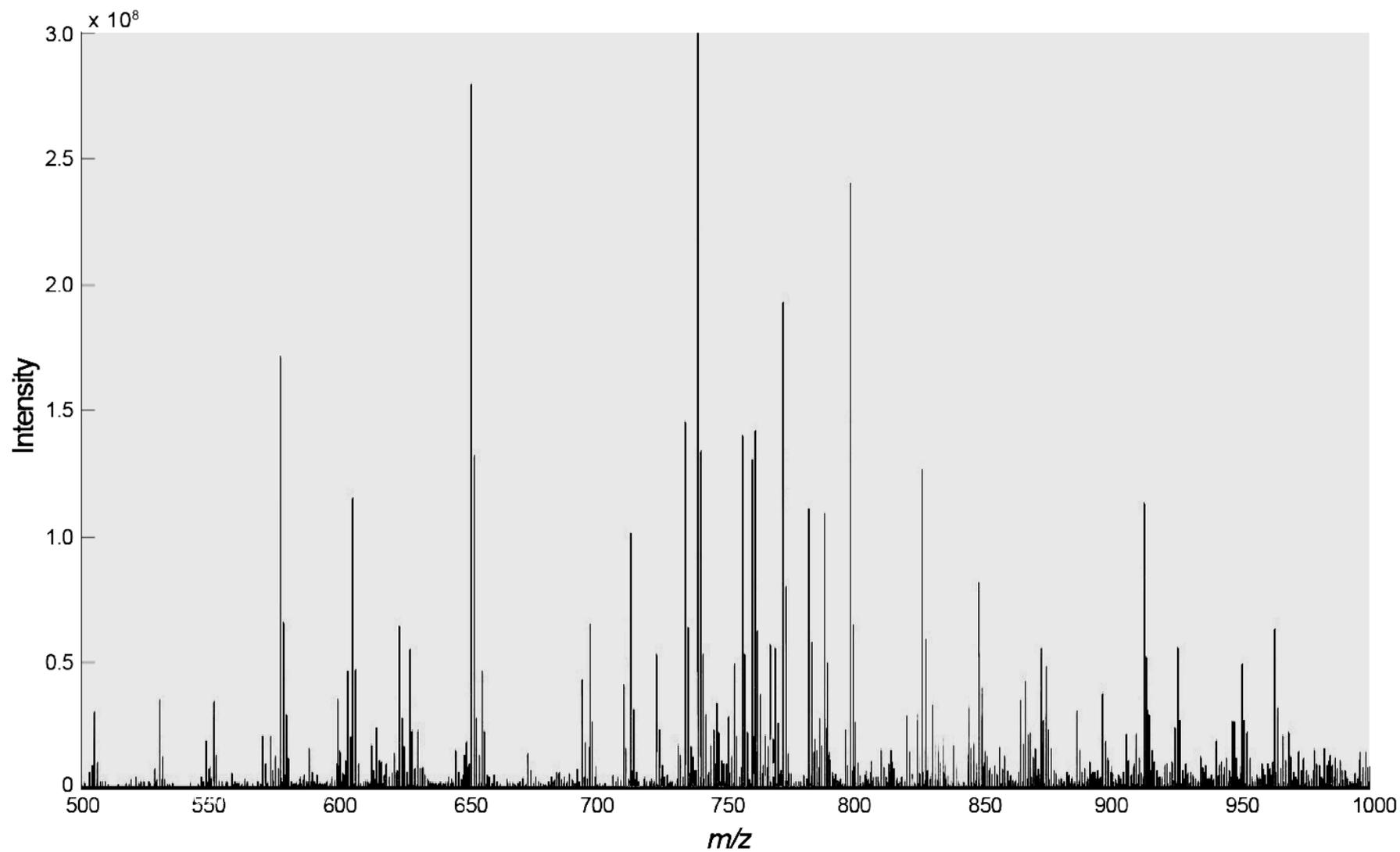
High Dynamic Range FT-ICR MS

- High Dynamic Range MALDI FT-ICR Example

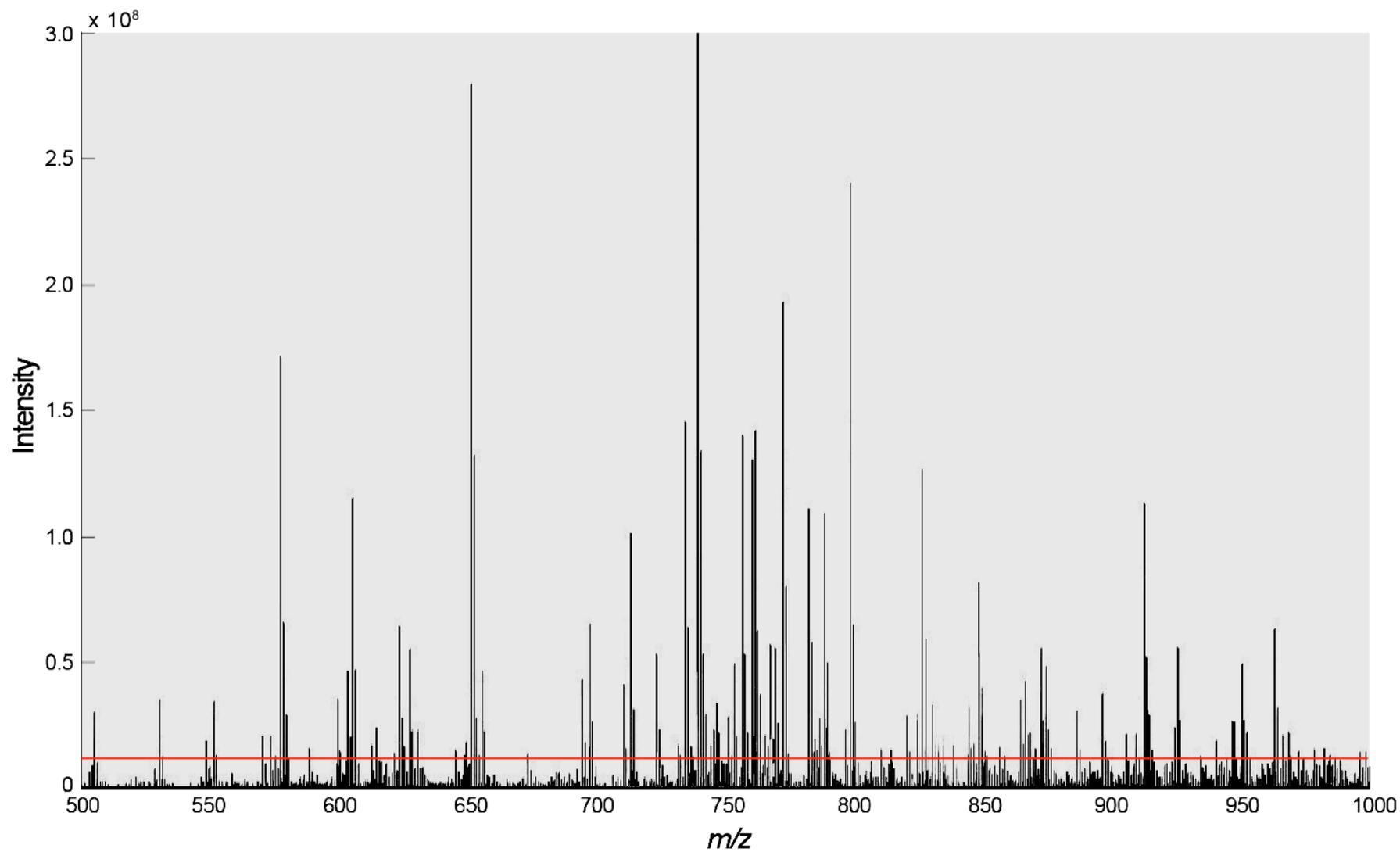
- Continuous Accumulation of Selected Ions (CASI)
- Result: 3 orders of magnitude increase in sensitivity.
- Process repeated to cover entire mass range.



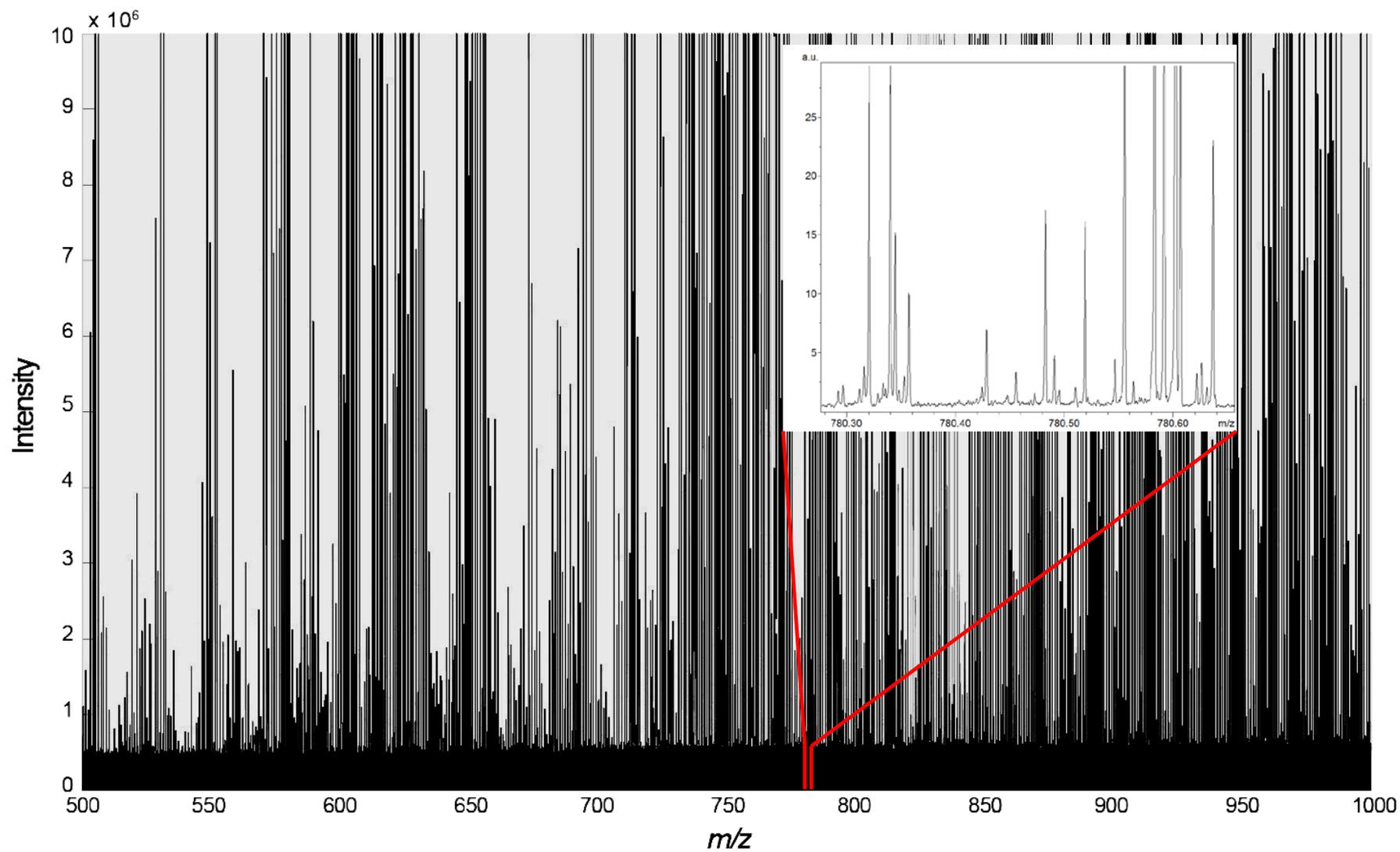
High Dynamic Range FT-ICR MS



High Dynamic Range FT-ICR MS



High Dynamic Range FT-ICR MS



Proof of Concept: MALDI FTICR MS lipid imaging

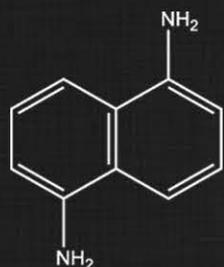
Sample Preparation

6 day old mouse pup

12 μm tissue section

Matrix: 1,5-Diaminonaphthalene

Matrix Application: Sublimation
(110 $^{\circ}\text{C}$, 50 mTorr, 6.75 min)



1,5-Diaminonaphthalene
(DAN)

Imaging MS Details

Bruker Solarix 9.4T FTICR MS

m/z 600 - 1600

256k TD (~ 0.5 s transient length)

ADD acquisition mode

50 μm spatial resolution

Laser focus: Small | ~ 45 μm

50 shots/px

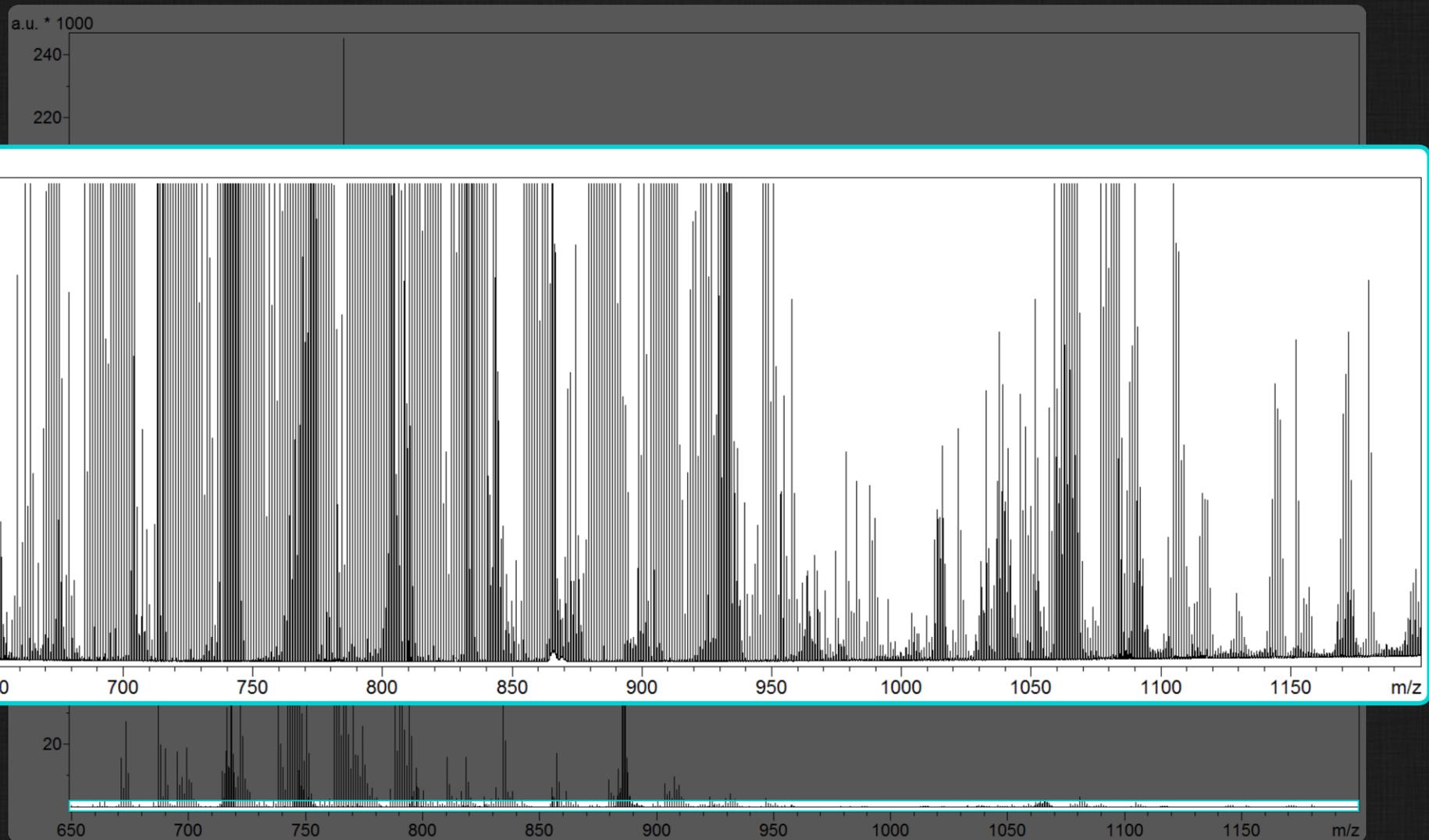
Laser Frequency: 2 kHz

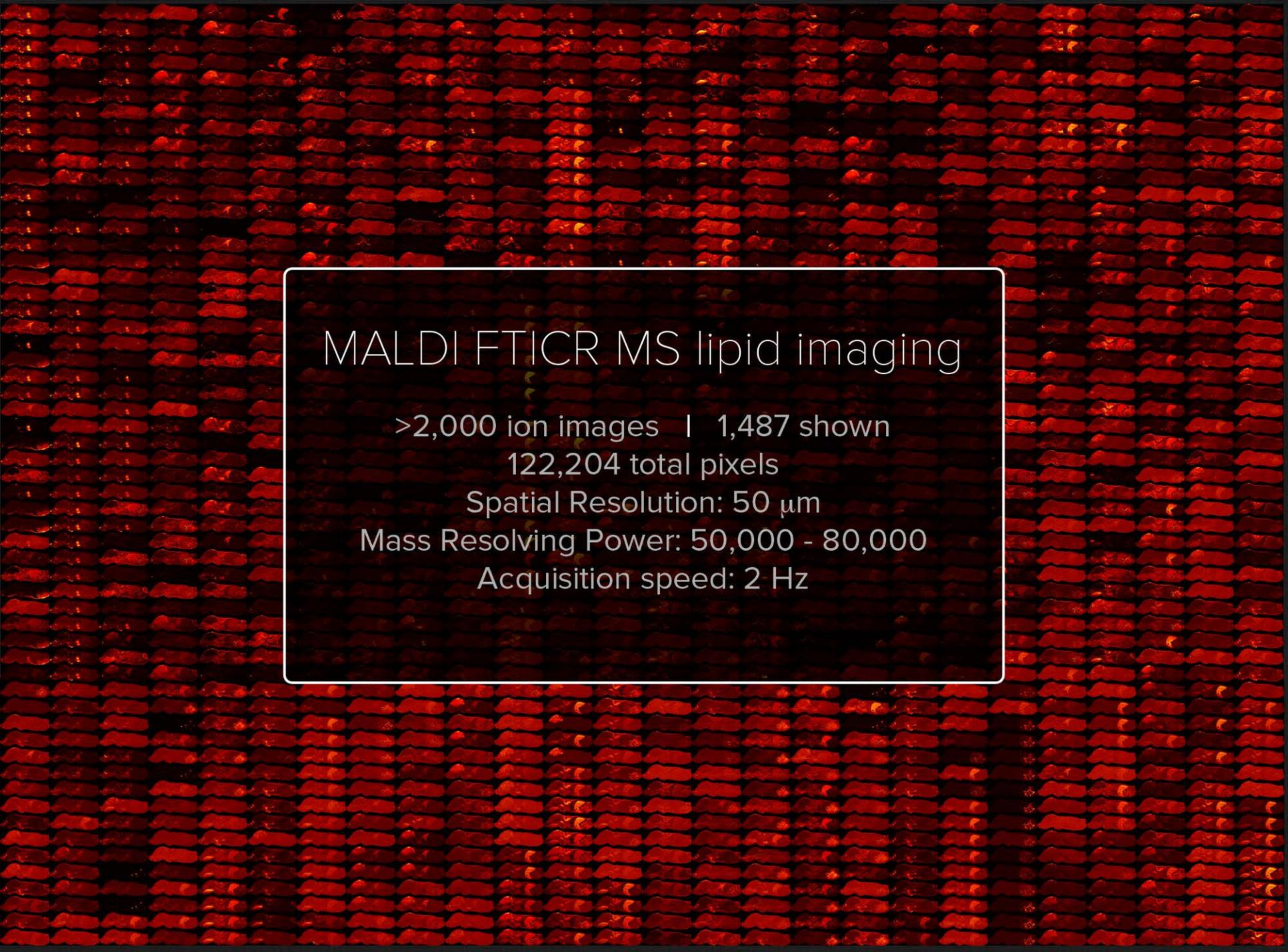
122,204 px



MALDI FTICR MS lipid imaging

average over entire image



The background of the slide is a dense grid of small, rectangular ion images. Each image shows a complex pattern of red and orange spots, representing the mass-to-charge ratio of ions detected at a specific spatial location. The overall appearance is a textured, multi-colored mosaic.

MALDI FTICR MS lipid imaging

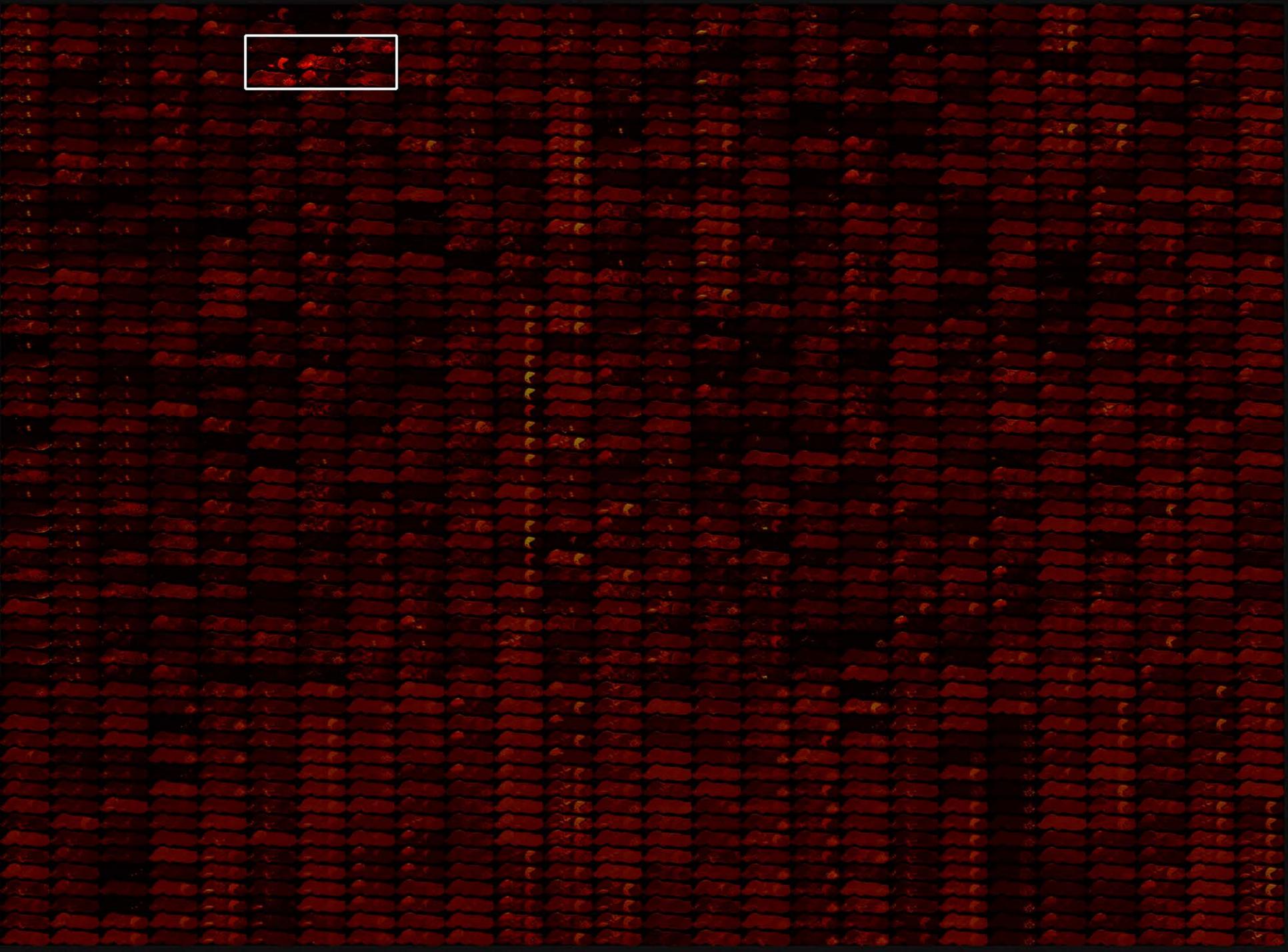
>2,000 ion images | 1,487 shown

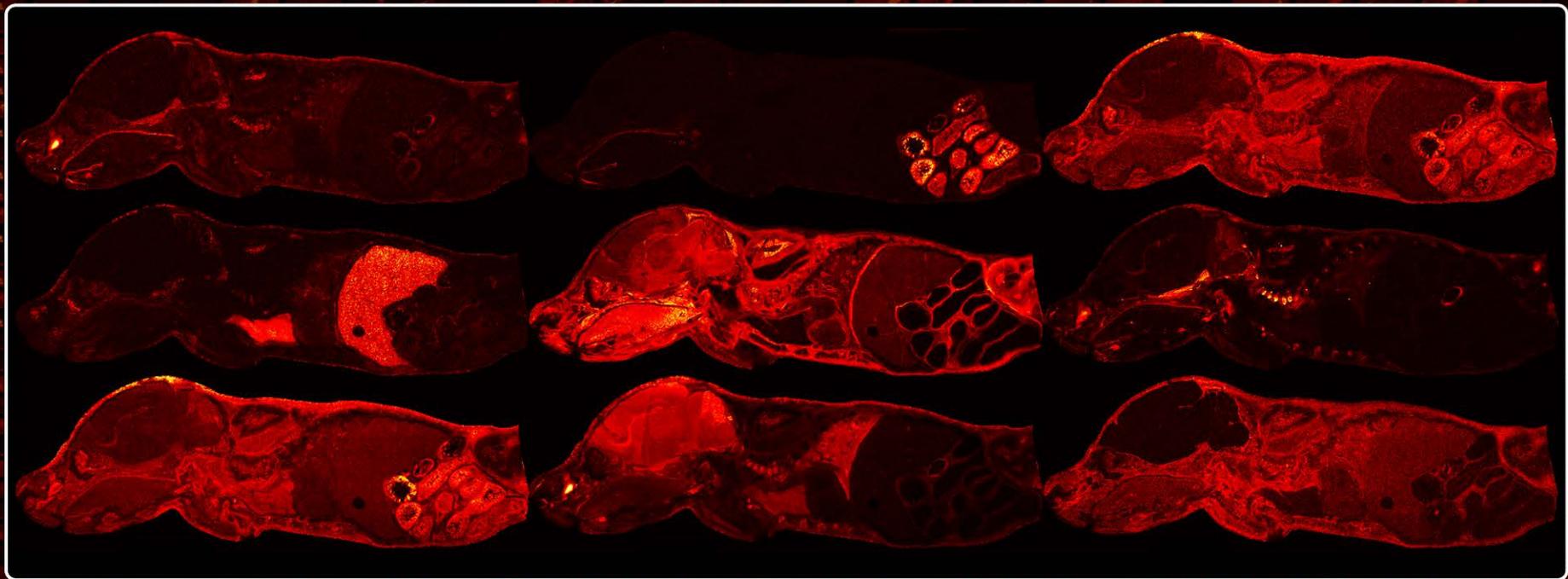
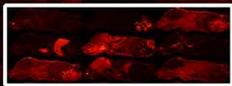
122,204 total pixels

Spatial Resolution: 50 μm

Mass Resolving Power: 50,000 - 80,000

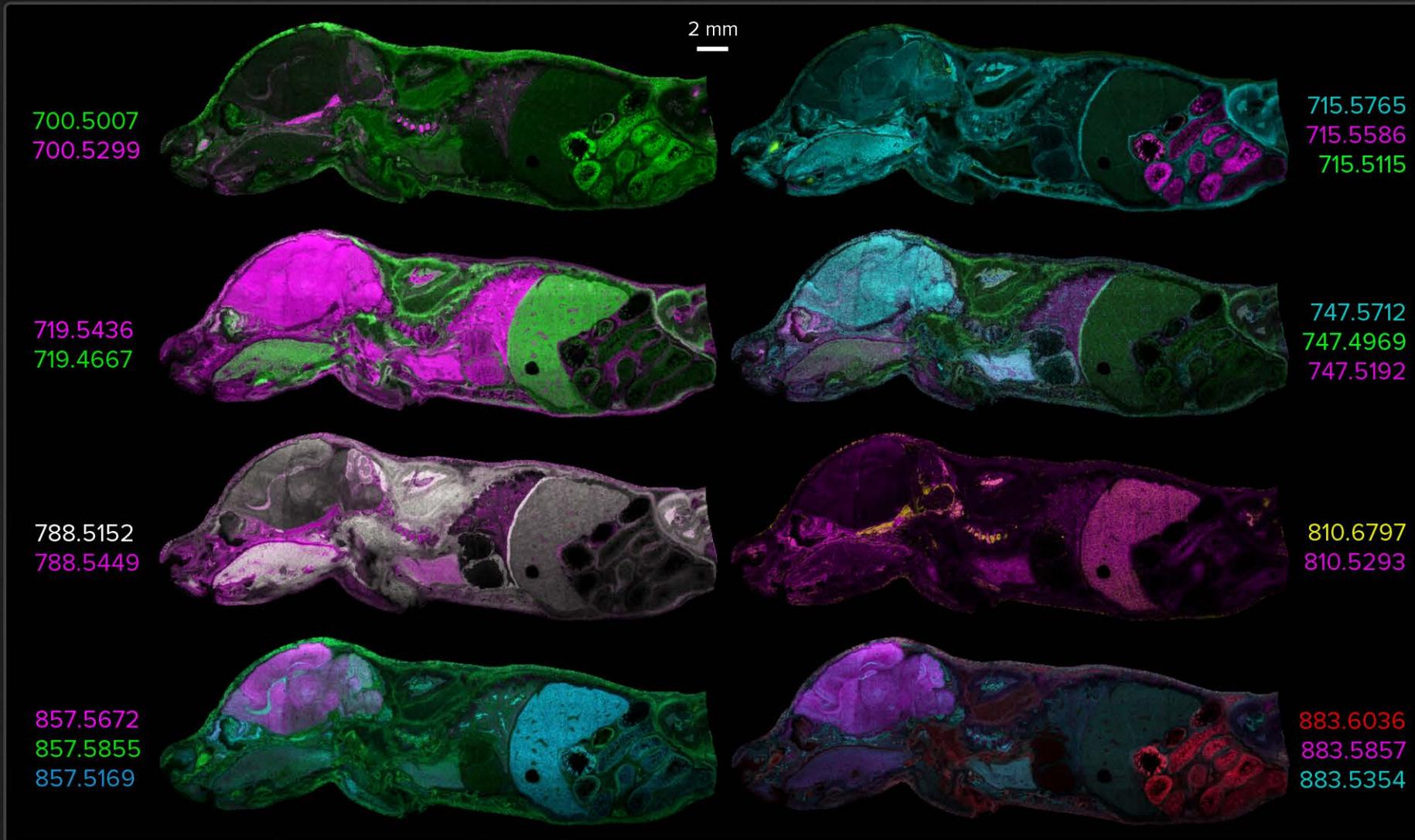
Acquisition speed: 2 Hz





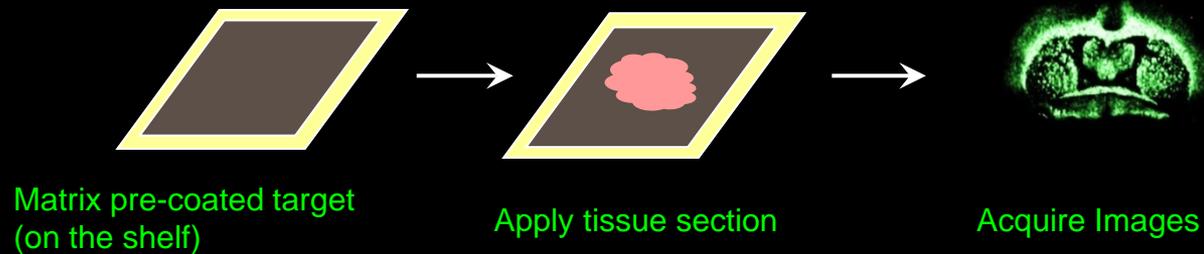
The Problem: IMS data complexity

Unique spatial distributions at each nominal mass



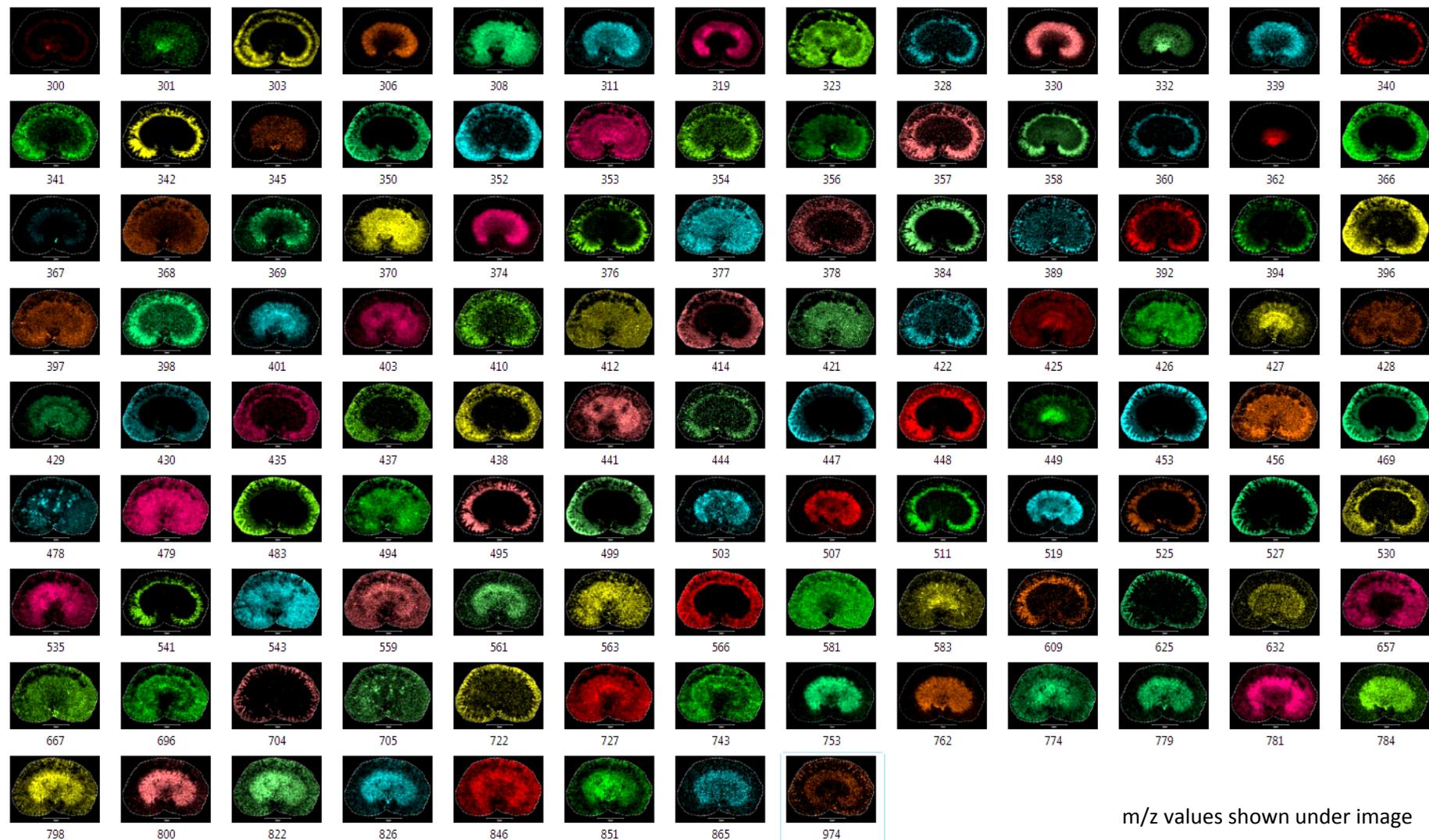
Improving Ease-of-Use

PRE-COATED TARGETS



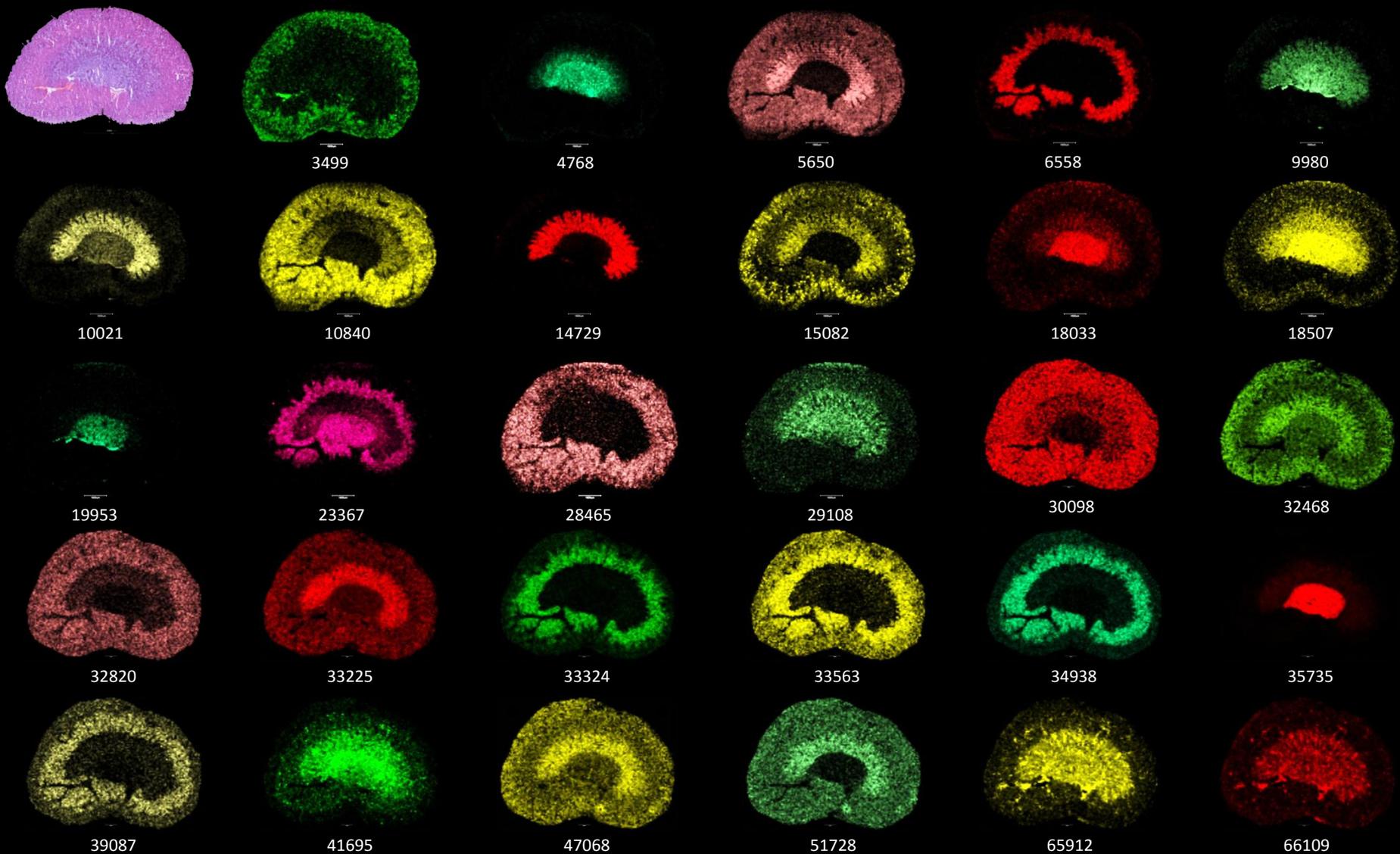
- Motivation:
 - Sample preparation is a perceived obstacle for the technology.
 - Users lack expertise
 - Time consuming
 - A matrix pre-coated target and an optimized protocol for use removes the burden of sample preparation from the end user.

Pre-coated CHCA for imaging lipids (positive mode)



m/z values shown under image

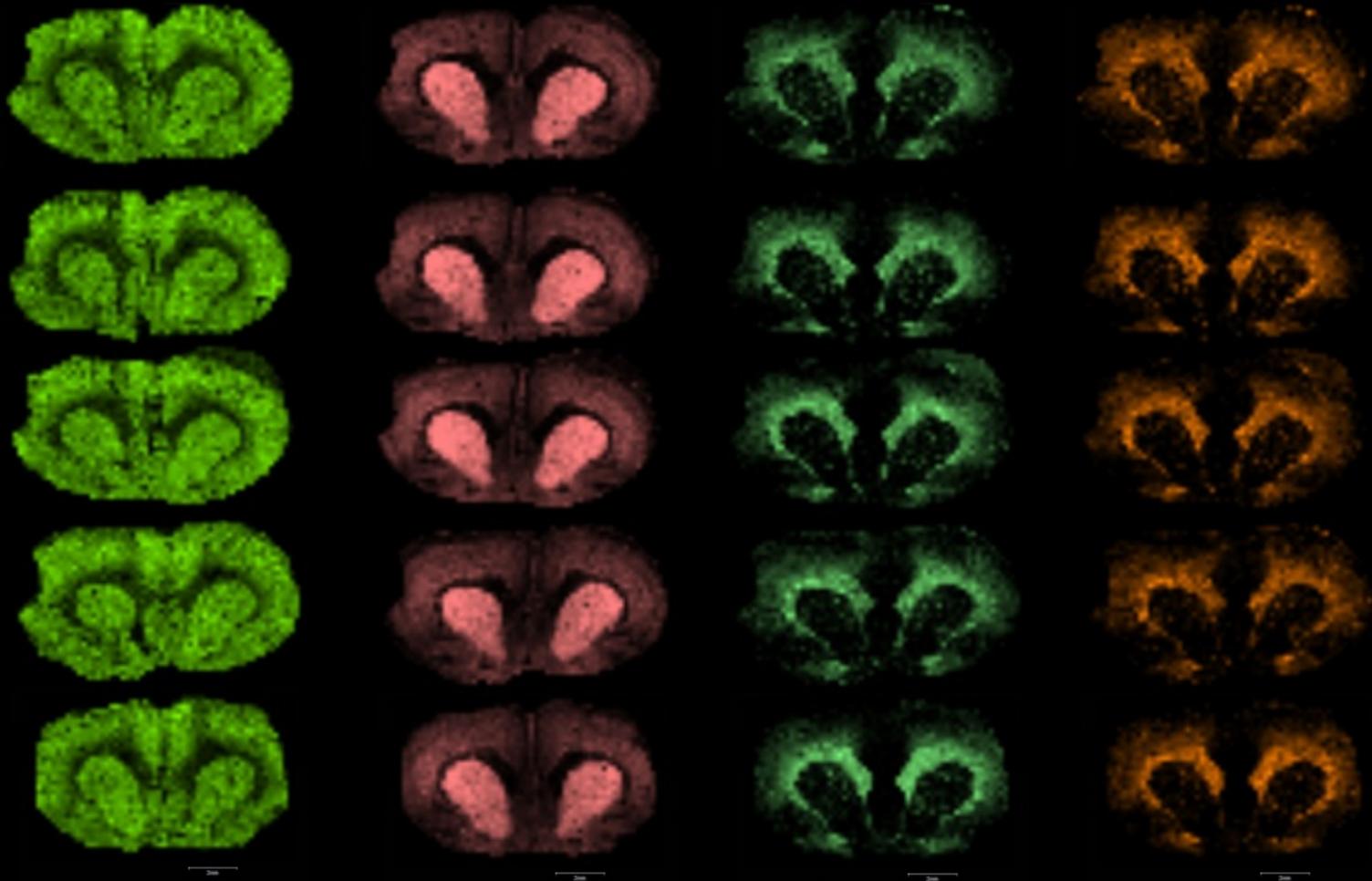
Pre-coated sinapinic acid for imaging proteins (3k to 70k Da)



m/z values shown under image

MATRIX PRE-COATED TARGETS PRODUCE REPRODUCIBLE IMAGES

MOUSE BRAIN ANALYZED USING A PRE-COATED SINAPINIC ACID TARGET



m/z 5628

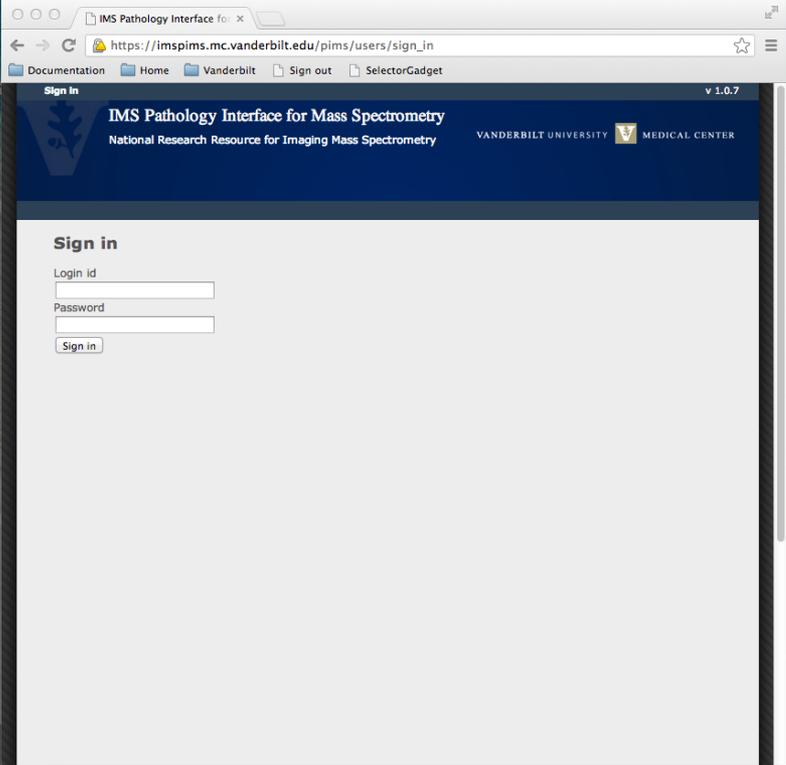
m/z 6710

m/z 7057

m/z 18,386

PATHOLOGY INTERFACE FOR MASS SPECTROMETRY (PIMS)

- Secure login provided to ensure access only to authorized users of the system.
- Access is controlled by the system administrator.
- Project level access is granted only to those collaborators involved in the project.
- Principal investigators and lab directors control access.

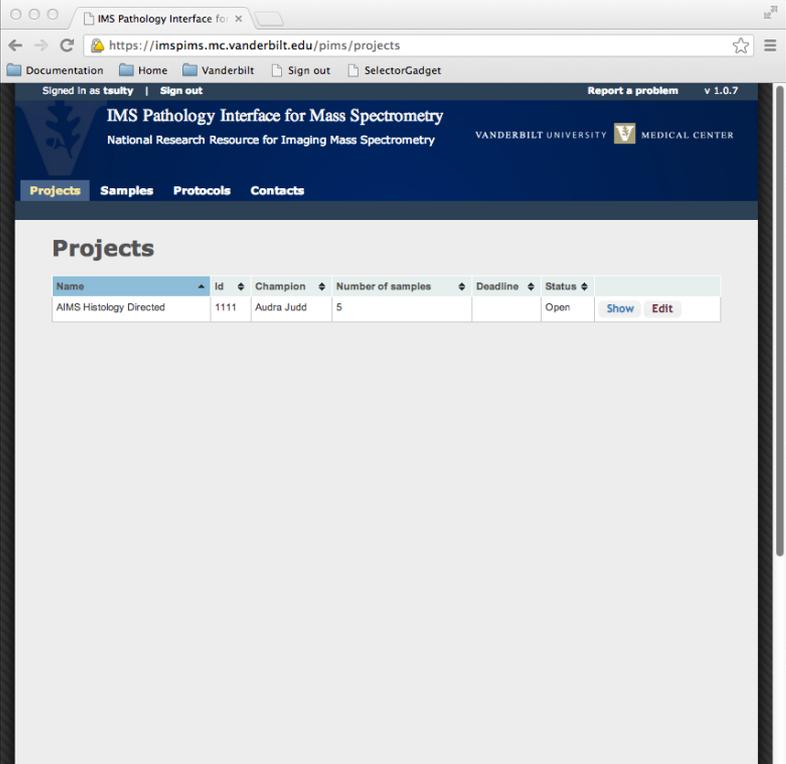


The screenshot shows a web browser window displaying the sign-in page for the IMS Pathology Interface for Mass Spectrometry (PIMS). The browser's address bar shows the URL https://imspims.mc.vanderbilt.edu/pims/users/sign_in. The page header includes the text "Sign in" and "v 1.0.7". The main header features the logo of the IMS Pathology Interface for Mass Spectrometry, the text "National Research Resource for Imaging Mass Spectrometry", and the Vanderbilt University Medical Center logo. The sign-in form contains the following elements:

- A "Sign in" heading.
- A "Login id" label followed by a text input field.
- A "Password" label followed by a text input field.
- A "Sign in" button.

PATHOLOGY INTERFACE FOR MASS SPECTROMETRY (PIMS)

- Database is organized by projects.
- All projects are shown with basic information about that project.
- Investigators can only view projects to which they have been assigned.



The screenshot displays the IMS Pathology Interface for Mass Spectrometry (PIMS) web application. The browser address bar shows the URL <https://imspims.mc.vanderbilt.edu/pims/projects>. The page header includes the text "Signed in as tsaulty | Sign out" and "Report a problem v 1.0.7". The main header features the IMS Pathology Interface for Mass Spectrometry logo and the text "National Research Resource for Imaging Mass Spectrometry" and "VANDERBILT UNIVERSITY MEDICAL CENTER". A navigation menu contains "Projects", "Samples", "Protocols", and "Contacts". The "Projects" section is active, displaying a table with the following data:

Name	Id	Champion	Number of samples	Deadline	Status	
AIMS Histology Directed	1111	Audra Judd	5		Open	Show Edit

PATHOLOGY INTERFACE FOR MASS SPECTROMETRY (PIMS)

- Samples assigned to projects are shown in the Project View.
- Collaborators input sample information for the study.
- Micrograph files are automatically assigned to samples based on the filename at import.
- Samples may be assigned to more than one project.

The screenshot displays the IMS Pathology Interface for Mass Spectrometry (PIMS) web application. The browser address bar shows the URL: <https://imspims.mc.vanderbilt.edu/pims/projects/6>. The page is signed in as 'tauity' and includes a 'Sign out' link. The main header identifies the application as the 'National Research Resource for Imaging Mass Spectrometry' at 'VANDERBILT UNIVERSITY MEDICAL CENTER'. The navigation menu includes 'Projects', 'Samples', 'Protocols', and 'Contacts'. The current view is 'Edit Project' for project ID 1111.

Project Information:

- Project Champion:** Audra Judd
- Name:** AIMS Histology Directed
- Project ID:** 1111
- Description:** Samples for AIMS workshop.
- Number of samples:** 5
- Deadline:**
- Budget Maximum:**
- Status:** Open

Groups:

Group Name	Samples in Group

Members:

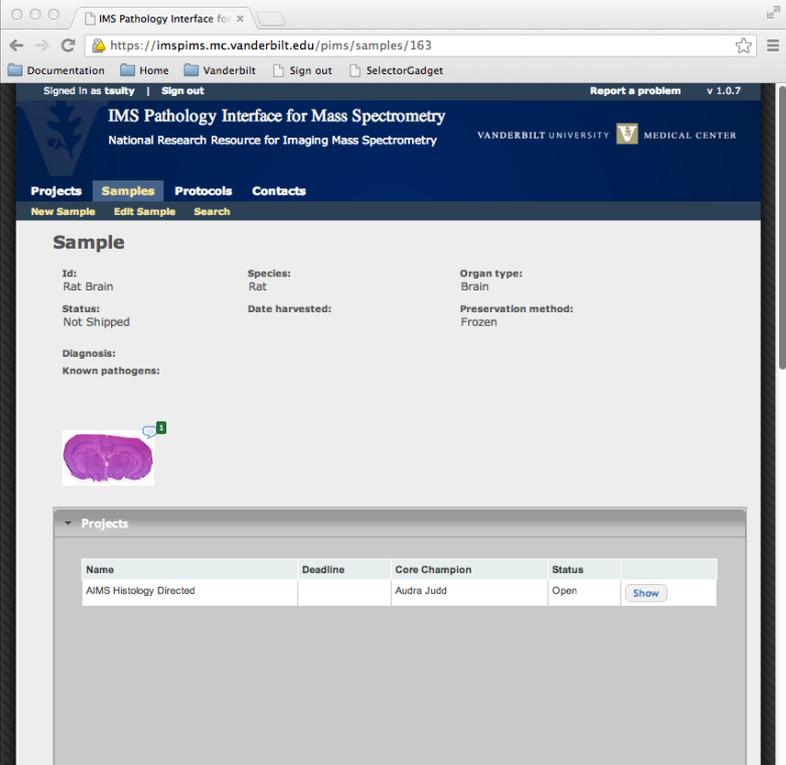
Name	Email	Project Role
Erin Seeley	erin.h.seeley@gmail.com	Project Manager
Tina Tsui	tsui12@gmail.com	Researcher

Samples Table:

Id	Species/Organ	Date harvested	Preservation method	Known pathogens	Last Image Upload	
Rat Brain	Rat/Brain		Frozen		2013/04/12 12:04:36	Show
Rat Brain 2	Rat/Brain		Frozen		2013/04/18 07:04:08	Show
Rat Heart	Rat/Heart		Frozen		2013/04/12 13:04:20	Show
Rat Kidney	Rat/Kidney		Frozen		2013/04/12 12:04:44	Show
Rat Kidney 2	Rat/Kidney		Frozen		2013/04/18 07:04:45	Show

PATHOLOGY INTERFACE FOR MASS SPECTROMETRY (PIMS)

- All sample information is displayed in Sample View along with thumb-nail images of the associated micrographs.
- All projects for which the sample has been assigned is shown in the dropdown below.
- Selection of thumbnails opens the image annotation window.



The screenshot displays the IMS Pathology Interface for Mass Spectrometry (PIMS) web application. The browser address bar shows the URL <https://imspims.mc.vanderbilt.edu/pims/samples/163>. The page header includes the text "Signed in as taulity | Sign out" and "Report a problem v 1.0.7". The main header features the logo for "IMS Pathology Interface for Mass Spectrometry" and "National Research Resource for Imaging Mass Spectrometry" at Vanderbilt University Medical Center. The navigation menu includes "Projects", "Samples", "Protocols", and "Contacts". Below the navigation, there are links for "New Sample", "Edit Sample", and "Search".

The "Sample" section displays the following information:

- Id:** Rat Brain
- Species:** Rat
- Organ type:** Brain
- Status:** Not Shipped
- Date harvested:**
- Preservation method:** Frozen
- Diagnosis:**
- Known pathogens:**

A thumbnail image of a brain section is shown below the sample information.

The "Projects" section is expanded to show a table of projects:

Name	Deadline	Core Champion	Status	
AIMS Histology Directed		Audra Judd	Open	Show

Sample ID

Annotation can be made for discrete spots (histology-directed) or by highlighting regions of interest (imaging).

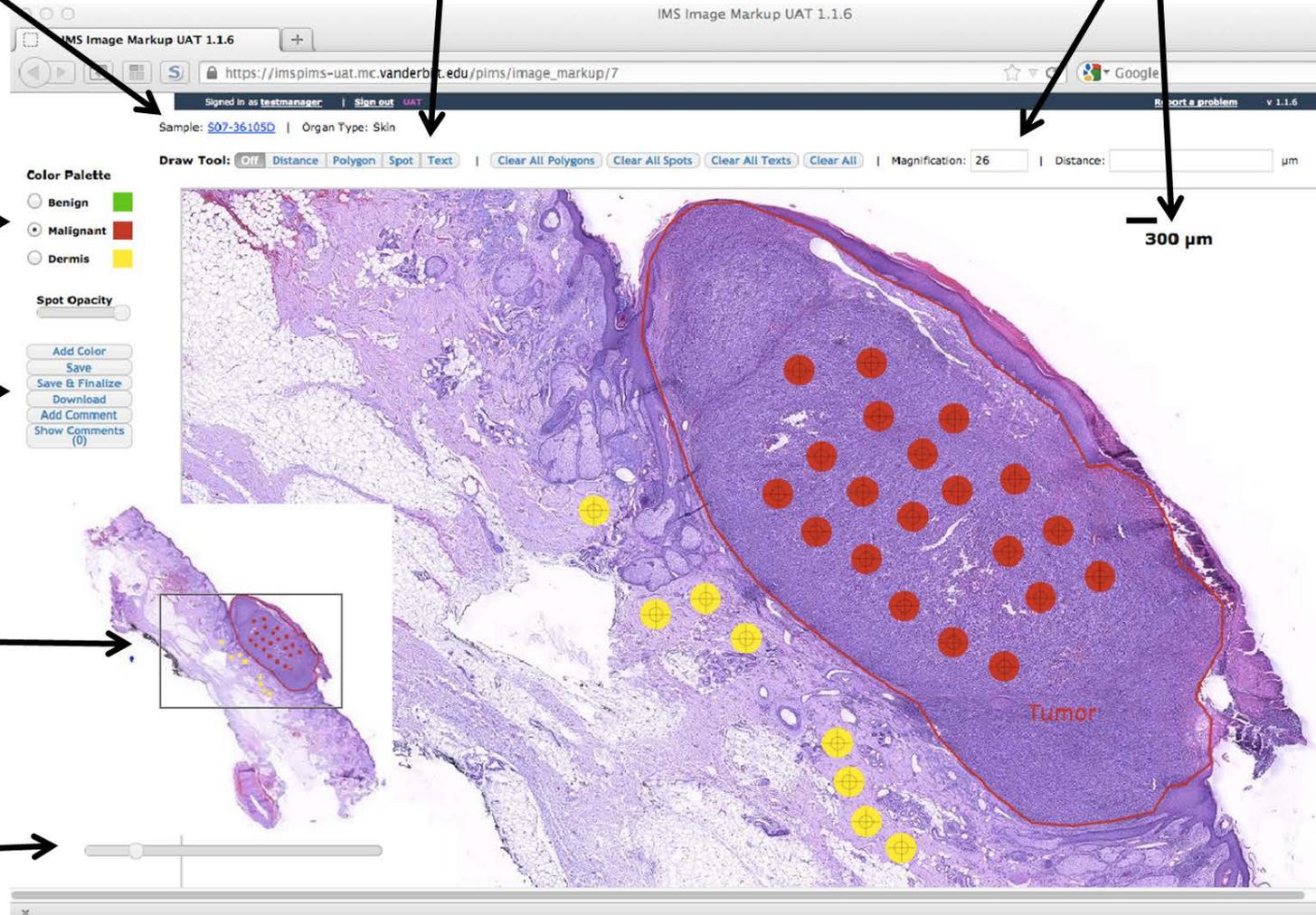
Scale bar and magnification provided/rescaled according to current zoom level.

User-defined color palette can be customized for each project.

Annotated images can be saved, exported and further documented using comments.

Navigation pane orients the user to the location within the sample.

Zoom feature allows the user to adjust the field of view.



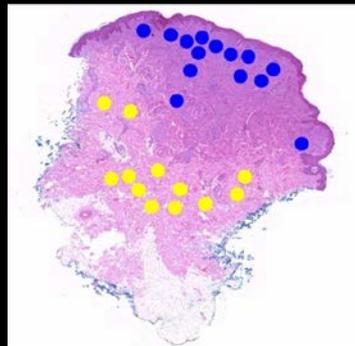
Case Study: Melanoma

- In 2012, an estimated 76,250 new cases were diagnosed (annual increase of 3% since 2004).
 - Source: American Cancer Society.
- The number of biopsies performed in the US to rule out melanoma range between 1-2 M per year. Of these, 25% cannot be definitively classified using routine histopathology.
 - Source: *Am J Surg Pathol*, 33(8), 1146-56.

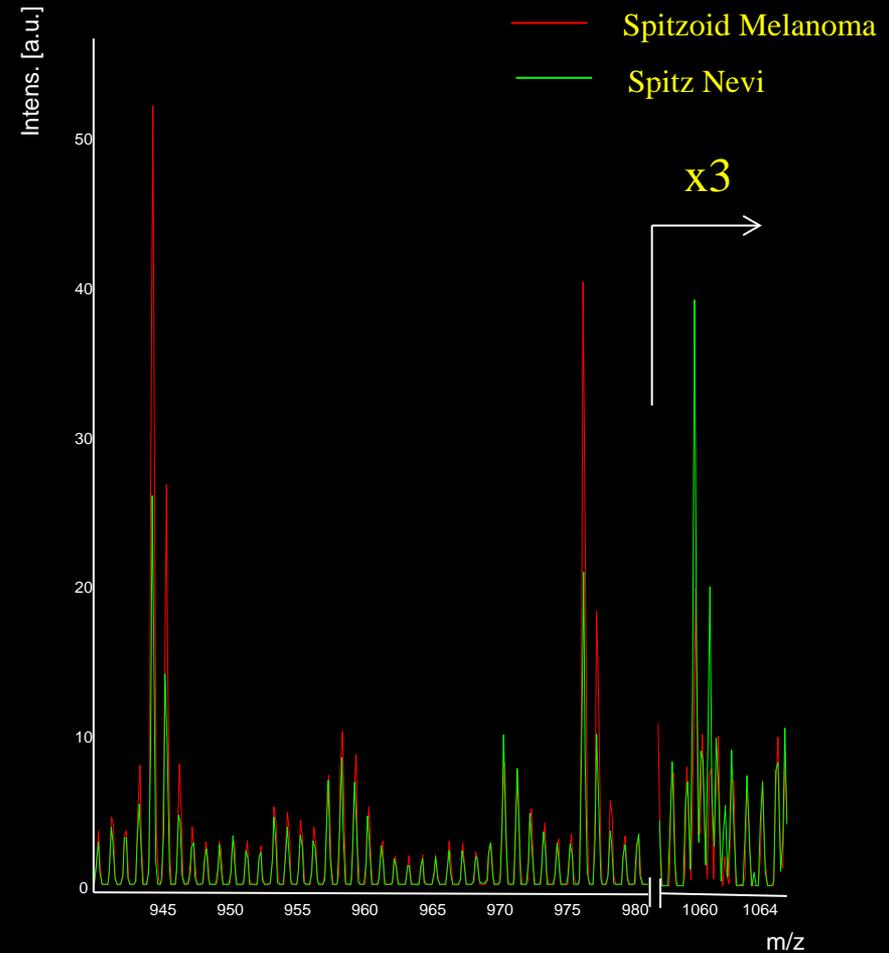
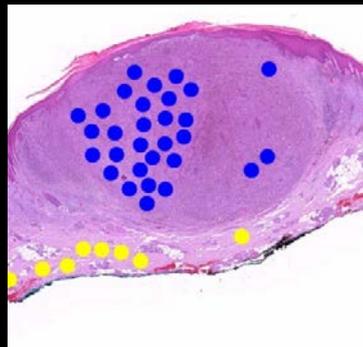
MS Analysis of Spitzoid Lesions in FFPE Biopsies

Lazova, R.; et al. *Am J Dermatopathol.* **34**, 82-90 (February 2012).

Spitz nevus



Spitzoid Melanoma



Classification of Spitzoid Lesions

56 SN and 54 SMM from Yale University Spitzoid Neoplasm Repository

Training set	# Patients	Classification Accuracy (%)
Spitz nevi (SN)	26	100
Spitzoid Malignant Melanoma (SMM)	25	96

Validation (test) set	# Patients	Classification Accuracy (%)
Spitz nevi (SN)	30	97
Spitzoid Malignant Melanoma (SMM)	29	90

International Spitzoid Neoplasm Study Group



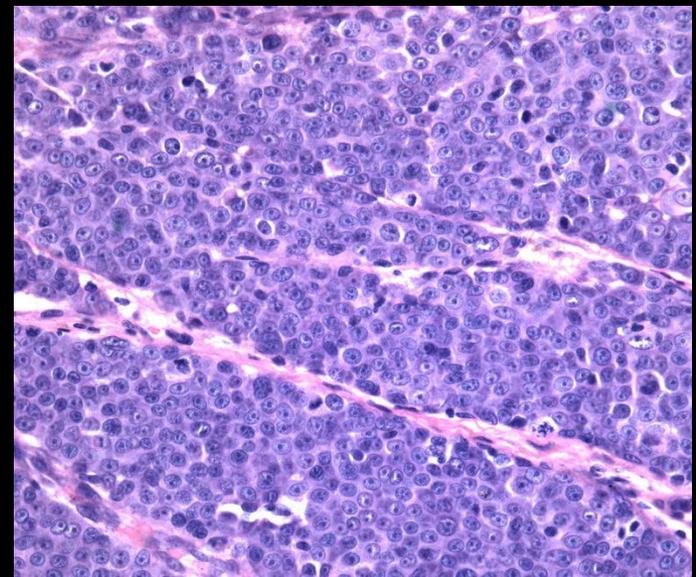
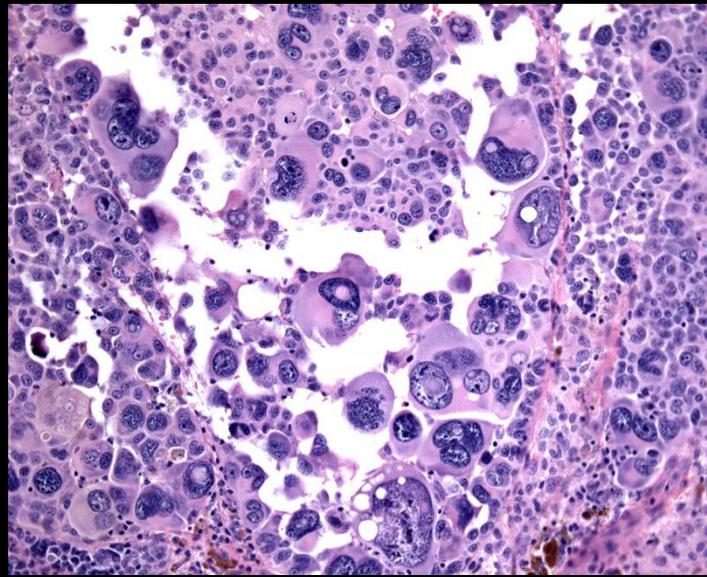
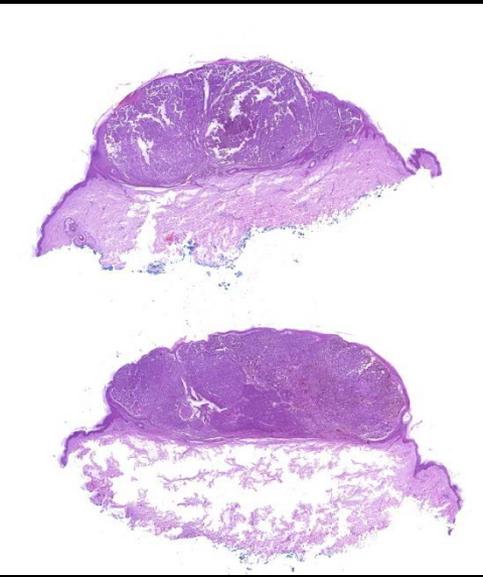
Atypical Spitzoid Neoplasms

#	Age	Gender	Site	Histologic Dx	MS Dx	Follow up (y)	Clinical Status
1	43	M	Back	SMM	SMM	3.5	Negative LN; ANED
2	23	F	L calf	SMM	SMM	2	Positive LN; ANED
3	28	F	Thigh	SMM	SMM	12	Positive LN 8 years later; ANED
4	6	F	L neck	SMM	SMM	1.5	Positive LN; ANED
5	39	F	L post leg	SMM	SMM	1.5	Positive LN; ANED
6	5	F	Buttock	SMM	SMM	6	Positive LN; ANED
7	29	F	R upper back	SMM	SMM	14	Negative LN: Re-excision; ANED;
8	50	M	thorax	SMM	SMM	3	DOD with lung mets 3 years later
9	43	M	back	SMM	SMM	4	Negative LN; ANED
10	57	F	NK	SMM	SMM	3	Negative LN; ANED
11	15	F	L neck	SMM	SN	4	Negative LN; ANED
12	6	M	Abdomen	SMM	SN	1	ANED
13	44	F	R upper arm	SMM	SN	7	ANED; 2 other ASN favor SN
14	16	M	Back	SMM	SN	10	Negative LN; ANED
15	55	M	R mid back	SMM	SN	2	ANED
16	40	F	R upper arm	SMM	SN	11	Negative LN; ANED
17	9	M	R upper arm	SMM	SN	14	Negative LN; ANED
18	17	M	Chest	SMM	SN	1	Negative LN; ANED
19	54	F	R upper arm	SMM	SN	8	Negative LN; ANED
20	44	F	R buttock	SMM	SN	9	Negative LN; ANED
			R upper arm	SMM	SN	8	Negative LN; ANED
21	30	F	R shin	SMM	SN	14	ANED
22	57	M	R thigh	SMM	SN	12	ANED
23	46	M	R arm	SMM	SN	4	1 Positive LN-1 cell; ANED
24	54	F	R upper arm	SMM	SN	8	Negative LN; ANED

LN – Lymph Node; ANED – Alive, No Evidence of Disease; DOD – Dead of Disease

Case Study

- 36 year old pregnant woman presents with lesion on upper arm
- Excisional biopsy performed and determined to be malignant
- Insufficient margins taken for size of lesion
- No further treatment during pregnancy

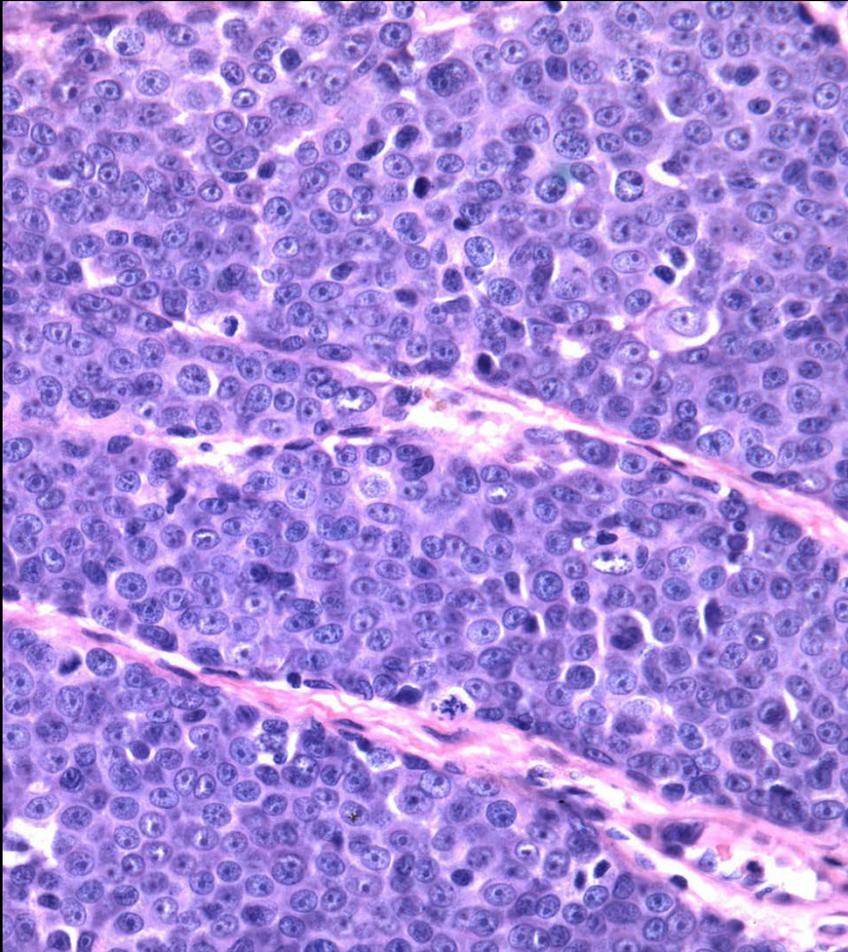


Case Study

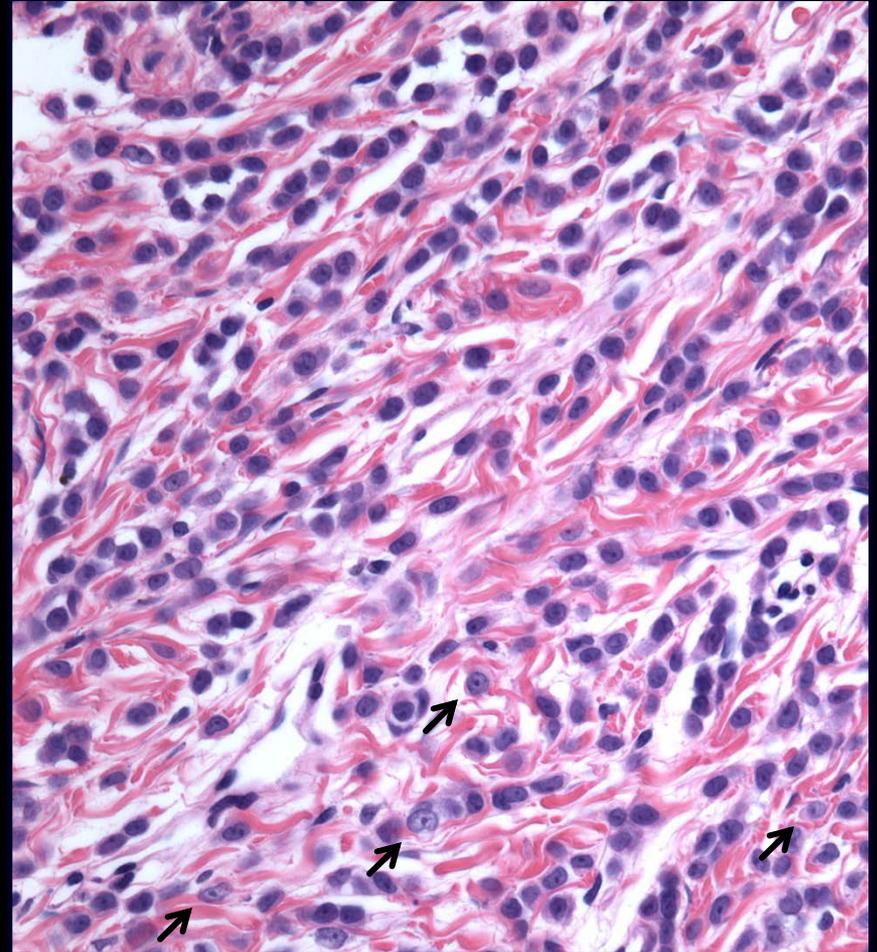
Two months later, male baby born with multiple nevi



Mother



Baby



Metastases or Congenital Nevi?

Mass Spectrometry Analysis

Mother

Mass Spectrometry

Clinical Diagnosis,
Malignant Melanoma

29/29 regions,
Malignant Melanoma

**Skin lesions
Baby**

Mass Spectrometry

Sample A: Indeterminate

9/9 regions,
Spitz nevus

Sample B: Indeterminate

23/23 regions,
Spitz nevus

Cells within lesions on baby contained y chromosome

Conclusions

- Evidence supports MALDI-based profiling as a useful clinical tool.
- Next-generation instrumentation is moving us closer to clinical applications of Imaging Mass Spectrometry.
 - Throughput
 - Spatial resolution
 - Expertise
- What's needed?
 - A more elegant solution.
 - Continue form build clinical case for tissue based classification.

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